



Australian Government



AUSTRALIAN INSTITUTE
OF MARINE SCIENCE

Coral Indicators for the 2025 Gladstone Harbour Report Card: ISP014

Cassandra Thompson, Angus Thompson, Johnston Davidson



Report Prepared for the Gladstone Healthy Harbour Partnership

AIMS: Australia's tropical marine research agency.

www.aims.gov.au

Australian Institute of Marine Science

PMB No 3
Townsville MC Qld 4810

PO Box 41775
Casuarina NT 0811

Indian Ocean Marine Research Centre
University of Western Australia, M096
Crawley WA 6009

This report should be cited as:

Thompson C, Thompson A, Davidson J (2025) Coral Indicators for the 2025 Gladstone Harbour Report Card: ISP014. Report prepared for Gladstone Healthy Harbour Partnership. Australian Institute of Marine Science, Townsville. (47 pp)

© Copyright: Australian Institute of Marine Science (AIMS) 2025

All rights are reserved, and no part of this document may be reproduced, stored or copied in any form or by any means whatsoever except with the prior written permission of AIMS

DISCLAIMER

While reasonable efforts have been made to ensure that the contents of this document are factually correct, AIMS does not make any representation or give any warranty regarding the accuracy, completeness, currency or suitability for any particular purpose of the information or statements contained in this document. To the extent permitted by law AIMS shall not be liable for any loss, damage, cost or expense that may be occasioned directly or indirectly through the use of or reliance on the contents of this document.

Project Leader shall ensure that documents have been fully checked and approved prior to submittal to client			
Revision History:	Name	Date	Comments
Prepared by:	Cassandra Thompson	22/07/2025	
Reviewed by:	Angus Thompson	23/07/2025	
Approved by:	David Wachenfeld	30/07/2025	
Reviewed by:	ISP	11/09/2025	
Revised by:	Cassandra Thompson and Angus Thompson	01/10/2025	Addressed comments from ISP review

Cover Photo:

A variety of common macroalgae fill the frame at Facing Reef: *Asparagopsis*, *Dictyota*, *Lobophora*, and *Sargassum*.
Photographer – Cassandra Thompson (AIMS).

CONTENTS

1	EXECUTIVE SUMMARY	1
2	BACKGROUND	3
3	METHODS.....	4
3.1	Sampling design.....	4
3.2	Survey methods.....	4
3.2.1	<i>Photo point intercept transects</i>	4
3.2.2	<i>Juvenile coral surveys</i>	4
3.2.3	<i>Scuba search transects</i>	5
3.3	Coral community sub-indicators	6
3.3.1	<i>Coral cover</i>	6
3.3.2	<i>Juvenile density</i>	6
3.3.3	<i>Macroalgae cover</i>	7
3.3.4	<i>Coral change</i>	7
3.3.5	<i>Scoring of sub-indicators</i>	8
3.3.6	<i>Aggregation of sub-indicator scores</i>	8
3.4	Key pressures.....	9
3.4.1	<i>Thermal bleaching</i>	9
3.4.2	<i>Runoff</i>	10
3.5	Plotting of results	10
3.5.1	<i>Time-series of indicator and sub-indicators</i>	10
3.5.2	<i>Comparison of Gladstone Harbour Coral communities to other near-shore reefs</i>	11
4	RESULTS.....	12
4.1	Coral cover.....	12
4.2	Macroalgae cover	14
4.3	Juvenile density	15
4.4	Change in hard coral cover	16
4.5	Environmental pressures.....	17
4.6	Comparisons with other near-shore reefs	19
5	CONCLUSION.....	23
6	ACKNOWLEDGMENTS.....	26
7	REFERENCES	26
8	APPENDICES	31

8.1	Appendix 1: Data Tables.....	31
8.2	Appendix 2: Rationale for sub-indicator selection and threshold setting.....	43
8.2.1	Coral cover	43
8.2.2	Macroalgae cover.....	44
8.2.3	Juvenile density	45
8.2.4	Cover change.....	46
8.2.5	References.....	48

TABLE OF FIGURES

Figure 1 Coral monitoring sites.....	5
Figure 2 Coral indicator for reporting zones.....	12
Figure 3 Trends in coral cover for reporting zones.	13
Figure 4 Composition of benthic cover at each location.	14
Figure 5 Trends in macroalgae cover for reporting zones.....	15
Figure 6 Trends in juvenile density for reporting zones.....	16
Figure 7 Composition of juvenile coral communities at each location.	16
Figure 8 Coral cover change timeseries.....	17
Figure 9 Weekly mean temperature profiles recorded at coral monitoring locations.....	18
Figure 10 Annual maximum Degree Heating Week estimates.	19
Figure 11 Ordination biplot comparing Gladstone Harbour reef communities with other near-shore reefs.....	20
Figure 12 Regional comparisons of sub-indicators.....	21

LIST OF TABLES

Table 1 Coral indicator and sub-indicator scores for 2025.....	1
Table 2 Thresholds and bounds for scoring coral sub-indicators.	8
Table 3 Conversion of aggregated indicator scores to report card grades.	9
Table Coral cover change sub-indicator scores for reefs and zones 2025.....	17
Table River discharge.	19
Table A 1 Site location and transect directions.....	31
Table A 2 Sub-indicator values for Gladstone Harbour.	32
Table A 3 Indicator and sub-indicator scores for Gladstone Harbour.	32
Table A 4 Sub-indicator values for reporting zones	33
Table A 5 Indicator and Sub-indicator scores for reporting zones.....	34
Table A 6 Sub-indicator values for individual reefs.....	35
Table A 7 Timeseries of indicator and sub-indicator scores for Mid Harbour reefs.....	37
Table A 8 Timeseries of indicator and sub-indicator scores for Outer Harbour reefs.	38
Table A 9 Genus level coral cover and abundance of juvenile corals at reefs surveyed in 2025.	39
Table A 10 Cover (%) of algae, sponges and sand and silt at reefs surveyed in 2025.	40
Table A 11 Causes of coral mortality at time of survey.....	41
Table A 12 Size-class distribution of juvenile corals.....	42

1 EXECUTIVE SUMMARY

This report presents a detailed description of the benthic communities at coral monitoring locations within the Mid and Outer Harbour reporting zones that form the basis of the coral community indicator of the 2025 Gladstone Harbour Report Card.

In April 2025, the Australian Institute of Marine Science (AIMS) resurveyed benthic communities at permanent coral monitoring locations in the Mid Harbour (four locations) and Outer Harbour (two locations). Overall, the condition of these communities remains 'very poor' and received a Grade E (Table 1, Figure 2).

Report card grades are based on the assessment of four indicators of reef condition: the proportion of the substrate occupied by living corals (Coral cover), the proportion of the substrate occupied by large fleshy species of algae (Macroalgae cover), the density of juvenile hard corals (Juvenile density) and the rate of change in coral cover relative to the expected change for a given community (Cover change).

Observed levels of the indicators were converted to scores based on thresholds developed for the 2018 Gladstone Harbour Report Card.

Table 1 Coral indicator and sub-indicator scores for 2025

Juvenile density	Coral cover	Macroalgae cover	Coral change	Coral indicator	
				Score	Grade
0.1	0.05	0.1	0.09	0.09	E

The 'very poor' grade for coral communities is heavily influenced by the continuing low cover of corals and high cover of macroalgae on most reefs. Mean coral cover (hard and soft coral combined) across Gladstone Harbour was 4%, down from the 6%, observed in the two preceding years and continues to be substantially lower than the 39% mean hard coral cover estimated in the Mid Harbour reporting zone in 2009 (BMT WBM 2013). Whilst the BMT WBM (2013) report does not provide a mean estimate for soft coral cover, data presented indicates that soft coral cover ranged between ~4% - 40%. Loss of coral cover coincided with major flooding in 2013 that almost certainly exposed corals to lethally low levels of salinity and high turbidity.

The current low cover of corals across the Harbour highlights the continued lack of recovery of coral communities over the last decade.

The sub-indicators, Macroalgae cover, Juvenile density, and Coral change, are included in the monitoring program to provide measures of the recovery potential of coral communities following the impact of acute events, such as the 2013 floods. All continue to predict limited recovery potential.

The cover of macroalgae remains high, which translates into the 'very poor' assessment for this sub-indicator. Macroalgae can limit coral recovery through a variety of pathways including direct competition for space and suppression of coral recruitment.

The 'very poor' assessment of the juvenile density sub-indicator is likely to reflect both the pressures imposed by high cover of macroalgae and limited availability of coral larvae due to low levels of broodstock within the Harbour and limited connectivity to sources that may exist outside the Harbour.

The score for the coral change sub-indicator has been reduced to 'very poor' for the first time since surveys began, driven by a substantial decline in the Outer Harbour score (Table 1, Table A 4). High water temperatures in early 2020 resulted in coral bleaching at the two Outer Harbour sites, Seal Rocks North and Seal Rocks South. In early 2024 high water temperatures again caused coral bleaching across the region, which was observed to be still ongoing during surveys in May. The stress incurred during these bleaching events likely compounded the ongoing pressure imposed by abundant macroalgae and will have contributed to the observed slow recovery of coral cover.

In early 2025 the discharge from both the Calliope and Fitzroy rivers was higher than the long-term average by 1.3 and 2.2 times respectively, which adds further stress to corals and hampers recovery (Table 5). The continued prevalence of bio-eroding sponges, that kill corals across the Harbour, further limits the recovery of these reefs.

The clear lack of recovery of coral communities over a decade of monitoring suggests that reefs within the harbour are in a state of hysteresis whereby the high cover of macroalgae relative to corals is being perpetuated. The active removal of macroalgae species common to Gladstone Harbour, and in particular *Sargassum* has been shown to increase both the settlement of coral larvae and growth of adult corals, proffering a potential avenue for coral reef restoration within the harbour.

2 BACKGROUND

Coral communities around the world are under increasing pressure as intensifying land use, urbanisation and industrial development impinge on corals' ability to resist, or recover from, natural disturbances such as floods and storms or coral bleaching. Along the Great Barrier Reef (GBR) coast it is well documented that loads of sediments, nutrients and chemical contaminants carried to the sea in catchment runoff have increased since European settlement (Kroon *et al.* 2012, Waters *et al.* 2014).

Within Gladstone Harbour, coral communities are subject to the same range of pressures as other inshore coral reefs in the GBR (e.g., Storm events including cyclones, freshwater inundation from floods, heat stress), compounded by additional pressures associated with the operations of the Harbour and associated industries (e.g., Increased turbidity). It is for this reason that AIMS has co-invested with the Gladstone Healthy Harbour Partnership (GHHP) to monitor and report the condition of coral communities within the GHHP reporting area as part of the Gladstone Harbour Report Card.

The indicators, sampling methodology, and scoring system used to derive grades for the Gladstone Harbour Report Card were chosen to be as compatible as practicable to those used for the Great Barrier Reef Report Card (Queensland Government 2015). We note that revisions of the methods used to score coral community condition for the Great Barrier Reef Report Card (Thompson *et al.* 2016) mean that while indicators remain the same, thresholds against which state and regional report card scores are derived now differ from those used for the Gladstone Harbour Report Card for the Macroalgae and Coral cover indicators. The scoring for the Juvenile density indicator for the Gladstone Harbour Report Card was realigned to the State and other Regional report cards in 2018.

This report presents the eleventh annual survey of permanent coral monitoring transects initiated in 2015. The purpose of this report is to provide a detailed description of reef communities as observed in 2025 that expands on the necessarily succinct summary of condition presented by the 2025 Gladstone Harbour Report Card.

3 METHODS

3.1 Sampling design

Coral communities are monitored along permanently marked transects. The selection of sites and construction of transects occurred in July 2015 as reported in detail in Thompson *et al.* (2015). In brief, suitable sites were identified at four locations within the Mid Harbour reporting zone and two locations in the Outer Harbour reporting zone (Figure 1). Within each site a series of five 20 metre-long transects, each separated by a space of 5 metres, were constructed along a depth contour identified as the most suitable coral habitat; depths ranged between 0 and 1 metre below lowest astronomic tide (Table A 1) as dictated by the limited depth of hard coral communities within the Harbour. To ensure accurate relocation of sampling, the start of each transect was marked with a steel star-picket, with additional transect markers consisting of lengths of 10 mm steel rod placed at the midpoint and end of each transect. The starting point of the 1st transect was recorded as a GPS location (WGS84 datum) and compass bearings recorded along each transect to aid future relocation (Table A 1). At each transect the following three surveys of the benthic communities are undertaken annually. This report presents data collected on the 19th of April 2025.

3.2 Survey methods

3.2.1 Photo point intercept transects

Estimates of the composition of benthic communities were derived from the identification of organisms on digital photographs taken along the permanently marked transects. The method closely followed Standard Operation Procedure Number 10 of the AIMS Long-Term Monitoring Program, with minor differences due to the use of shorter transects on these smaller inshore reefs (LTMP, Jonker *et al.* 2020). The methods mirror those used by the [Marine Monitoring Program](#) (MMP). Digital photographs were taken at 50 cm intervals along each transect. Estimations of proportional cover of benthic community components were derived from the identification of the benthos lying beneath five fixed points digitally overlaid onto these images. Benthic cover of any group of interest is estimated as the proportion of all points that were identified and categorised as that group. A total of 32 images were analysed from each transect. Hard and soft corals were identified to genus level. Identifications for each point were entered directly into a data entry front-end to an Oracle® database, developed by AIMS. This system allows the recall of stored transect images and checking of all identified points.

3.2.2 Juvenile coral surveys

The number of juvenile coral colonies were counted *in situ* along the permanently marked transects. Prior to 2018, corals in the size classes: 0-2 cm, >2-5 cm, and >5-10 cm found within a strip 34 cm wide (data slate length) positioned on the upslope side of the transect line were identified to genus level. Since 2018, reporting of the >5-10 cm size class was discontinued, aligning the methodology used here with that used by the MMP (Thompson *et al.* 2016). Importantly, this method aims to record only those small colonies assessed as juveniles, i.e. which result from the settlement and subsequent

survival and growth of coral larvae, and so excludes small coral colonies considered to have resulted from the fragmentation or partial mortality of larger colonies.

Limiting observations to <5 cm more accurately focuses on juvenile rather than fragmented colonies and helps to exclude small colonies of slow growing corals which do not reflect the recent recruitment and survivorship dynamics which are assessed by this indicator. Further, the realignment of methodology allows direct comparison between Gladstone Harbour coral communities and those of other inshore reefs monitored by the MMP.

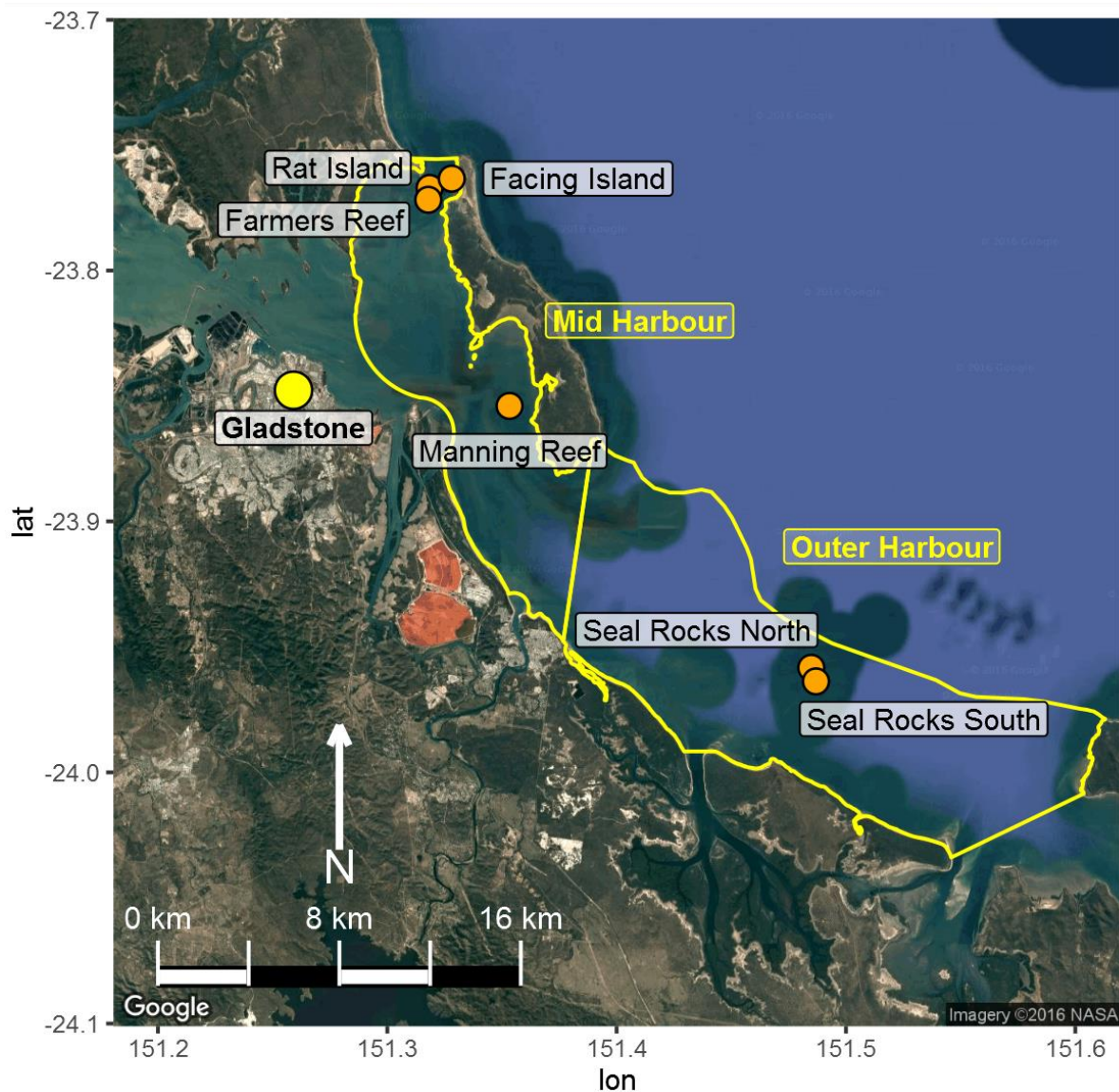


Figure 1 Coral monitoring sites.

3.2.3 Scuba search transects

Scuba search transects document the incidence of disease and other agents of coral mortality observed at the time of survey. This method closely followed the Standard Operation Procedure Number 9 of the AIMS Long-Term Monitoring Program (Miller *et al.* 2009) and serves to help identify probable causes of declines in coral community condition. For each 20 metre-long transect, a search was conducted within a 2 metre-wide belt centred on the marked transect line and the incidence of:

coral disease, coral bleaching, coral predation by *Drupella* or crown-of-thorns seastars, overgrowth by sponges, smothering by sediments or physical damage to coral colonies was recorded.

3.3 Coral community sub-indicators

The coral index is formulated around the concept of community resilience. The underlying assumption is that a 'resilient' community should show clear signs of recovery after inevitable acute disturbances, such as cyclones and coral bleaching events, or, in the absence of disturbance, maintain a high cover of corals and successful recruitment processes. For the Gladstone Harbour Report Card four sub-indicators of coral communities are included, each representing different processes that contribute to coral community resilience.

This section provides an overview of the methods used to estimate and score each sub-indicator that, in combination, capture both the state and resilience of coral communities. A full description for the rationale behind the selection and scoring of each sub-indicator is included in Appendix 2.

3.3.1 Coral cover

The most tangible and desirable indication of a healthy coral community is an abundance of coral. The Coral cover sub-indicator score at each reef is based on the proportional area of substrate covered by either 'Hard' (order Scleractinia) or 'Soft' (subclass Octocorallia) corals.

$$\text{coral cover}_{ij} = HC_{ij} + SC_{ij}$$

Where, *HC* and *SC* are the proportion of benthos occupied by hard and soft corals respectively,

i = reef and *j* = time.

While high coral cover generally indicates conditions that support coral growth and survival, low cover on its own does not always indicate the opposite, as reefs may be in a recovery phase after disturbance. However, if coral cover remains low over extended periods (e.g., a decade), this more strongly suggests that environmental conditions are limiting coral recovery. Coral communities are naturally dynamic being impacted by acute disturbance events such as cyclones, temperature anomalies and, in coastal areas, flooding. The sub-indicators: juvenile density, macroalgae cover, and coral change, were included as they represent the potential for coral communities to recover from disturbances.

3.3.2 Juvenile density

The density of juvenile corals is an indicator of the successful completion of early life history stages of corals from gametogenesis through fertilisation, larval survival in the plankton, settlement to the substrate and then early post settlement survival, all of which may be impacted by poor water quality (reviewed by Fabricius 2005, van Dam *et al.* 2011, Erftemeijer *et al.* 2012). The juvenile density sub-indicator was derived from counts of juvenile hard corals along belt transects and converted to a density per area of potentially colonisable hard substrate; estimated as the proportion of benthos identified as algae along the co-located point intercept transects:

$$\text{juvenile density}_{ij} = J_{ij} / AS_{ij}$$

Where, *J*=count of juvenile colonies < 5 cm in diameter, *AS*=area of transect occupied by algae, *i*=reef and *j*=time.

3.3.3 Macroalgae cover

High macroalgal abundance may suppress the recovery of coral communities through a variety of mechanisms ranging from competition with surviving colonies through to suppression of the recruitment process (e.g., McCook *et al.* 2014, Hughes *et al.* 2007, Foster *et al.* 2008, Cheal *et al.* 2013, Hauri *et al.* 2010). The values of the macroalgae cover sub-indicator were estimated as the proportion of benthos along point intercept transects identified as macroalgae:

$$\text{macroalgae cover}_{ij} = MA_{ij}$$

Where, MA is the proportion of the benthos occupied by macroalgae, i =reef and j =time. Macroalgae is here considered to include all algae larger than the filamentous turf or crustose coralline forms.

3.3.4 Coral change

While high coral cover can justifiably be considered a positive indicator of community condition, the reverse is not necessarily true. Low cover may occur following acute disturbance and, hence, may not be a direct reflection of the community's resilience to underlying environmental conditions. For this reason, in addition to considering the actual level of coral cover we assess the rate at which hard coral cover increases as a measure of recovery potential. The assessment of rates of cover increase is possible as rates of change in hard coral cover on inshore reefs have been modelled (Thompson *et al.* 2016), allowing estimations of expected increases in cover for communities of varying composition to be compared against observed changes.

A Bayesian framework was used to permit propagation of uncertainty through predictions of expected hard coral cover increase from separate models applied to fast growing corals of the family Acroporidae, and the combined cover of all other hard corals. Note that the example presented below for Acroporidae (Acr), has the same form as that applied for Other Corals ($OthC$) if these terms are exchanged where they appear in the equations.

$$\ln(Acr_{it}) \sim \mathcal{N}(\mu_{it}, \sigma^2)$$

$$\mu_{it} = vAcr_i + \ln(Acr_{it-1}) + \left(-\frac{vAcr_i}{\ln(estK_i)} \right) * \ln(Acr_{it-1} + OthC_{it-1} + Sc_{it-1})$$

$$vAcr_i = \alpha + \sum_{j=0}^J \beta_j Reef_i$$

$$\alpha \sim \mathcal{N}(0, 10^6)$$

$$\beta_j \sim \mathcal{N}(0, \sigma_{Reef}^2)$$

$$\sigma^2, \sigma_{Reef}^2 = \mathcal{U}(0, 100)$$

$$rAcr = \bar{vAcr}_i$$

Where, Acr_{it} , $OthC_{it}$ and Sc_{it} are the cover of Acroporidae coral, other hard coral and soft coral respectively at a given reef at time (t). $eskK$ is the community size at equilibrium (100-proportion of area comprised of unconsolidated substrates) and $rAcr$ is the rate of increase (growth rate) in cover of Acroporidae. Varying effects of Reef (β_j) is also incorporated to account for spatial autocorrelation. Model coefficients associated with the intercept, and Reef (α_i and β_j) all had weakly informative Gaussian priors (the latter two with model standard deviation). The overall rate of coral growth parameters ($rAcr$ or alternatively $rOthC$) constituted the mean of the individual posterior rates of increase ($vAcr_i$ or alternatively $vOthC_i$).

3.3.5 Scoring of sub-indicators

To facilitate the reporting of coral community condition the observed values for each sub-indicator were converted to scores on a common scale of 0 to 1. For each sub-indicator, observed values were scaled against thresholds which were set based on expert opinion and knowledge gained from the time-series of coral community condition collected by the Marine Monitoring Program (MMP) and the AIMS Long Term Monitoring Program (LTMP). Thresholds represent the boundary between report card grades of C and D (score = 0.5) that would indicate the switch between a community in satisfactory condition and one displaying a lack of resilience (Table 2). In addition, upper bounds were set that represent values of indicators that were considered to represent communities in as good a condition as could be expected in the local environment. Conversely, lower bounds were set to represent minimal resilience (Table 2). While observations may exceed these limits, any such values will be capped at the minimum or maximum score (0 or 1 respectively). For the cover change indicator, scores are averaged over three years of estimates excluding years during which reefs were categorised as having been impacted by an acute disturbance event. This averaging is done as expected increase in cover from one year to the next can be low, especially when coral cover is low, or the community is dominated by slow growing taxa. This averaging helps to account for sampling error inherent in each annual estimate of coral cover.

Table 2 Thresholds and bounds for scoring coral sub-indicators. Note that the thresholds for the juvenile density were updated in 2018 to account for the change in the methodology described above and are consistent with those used by the MMP on inshore reefs.

Indicator	Threshold (score = 0.5)	Upper bound (score = 1)	Lower bound (score = 0)
Coral cover	40%	90%	0%
Macroalgae cover	14%	5%	20%
Juvenile density	4.6 m ⁻²	13 m ⁻²	0 m ⁻²
Coral change	Lower 95% CI	2* upper 95% CI	Below 2* lower 95% CI

3.3.6 Aggregation of sub-indicator scores

The scaling of all scores to the common range of 0 to 1 allows aggregation of scores across sub-indicators at a hierarchy of spatial scales. Within this report, scores are presented at the scale of individual sub-indicators at each reef, individual sub-indicators and coral indicator scores for each reporting zone and the whole-of-harbour. Zone scores represent the mean score for each sub-indicator across reefs within that zone, while coral indicator scores represent the mean of the four zone-level sub-indicator scores. Similarly, harbour-wide scores were taken as the mean of the zone-level means for each sub-indicator and the coral indicator score as the mean of these harbour-wide sub-indicator scores. All scores were supplied to AIMS by the Data Integration and Management System based on sub-indicator values uploaded to that system by AIMS.

For the Gladstone Harbour Report Card, indicator scores are derived through the aggregation of bootstrapped distributions of sub-indicator scores, where bootstrapped distributions are produced by repeatedly sampling, with replacement, the observed distribution of sub-indicators. This method of aggregating distributions ensures that each distribution has equal weighting on the aggregation.

In practice, to aggregate sub-indicator scores at each reef to a mean score and estimate of variance for a zone requires that:

1. A bootstrap distribution of 10,000 samples is constructed for each sub-indicator within the zone.
2. The resulting bootstrap distributions are added together, and the mean indicator score for the zone along with variance extracted from this combined distribution.

Whole-of-harbour scores were similarly generated by respectively aggregating the sub-indicator distributions within zones, adding the aggregated distributions from each zone together to derive a harbour-level distribution from which mean and variance for sub-indicators at the scale of the Harbour were derived. Finally, adding the whole-of-harbour distributions for each sub-indicator yields the distribution from which the whole-of-harbour coral indicator score, and variance were extracted. Reef level coral indicator scores are simply the arithmetic mean of the scores for each sub-indicator.

Grades for coral community condition were derived from the scores estimated above according to the conversions described in Table 3.

Table 3 Conversion of aggregated indicator scores to report card grades.

A	Very good (0.85 - 1.00)
B	Good (0.65 - 0.84)
C	Satisfactory (0.50 - 0.64)
D	Poor (0.25 - 0.49)
E	Very poor (0.00 - 0.24)

3.4 Key pressures

Coral communities are susceptible to a range of pressures. Identifying these pressures and the associated drivers is essential in determining the likely cause of impacts to coral community condition. For inshore reefs of the GBR, common disturbances to coral communities include: physical damage caused by tropical cyclones (Osborne *et al.* 2011, De'ath *et al.* 2012), exposure to low salinity waters during flood events (van Woesik 1991, Jones & Berkelmans 2014), and anomalously high summer temperatures resulting in coral bleaching (Berkelmans *et al.* 2004, Sweatman *et al.* 2007). It is only once the influences of acute pressures have been accounted for that the potential impacts of chronic pressures such as elevated turbidity and nutrient levels can be inferred.

3.4.1 Thermal bleaching

Thermal stress, resulting in coral bleaching, is an increasing threat to coral communities in a warming world (Schleussner *et al.* 2016). During coral surveys in 2016 AIMS deployed temperature loggers to the pickets marking the first transect at each of Rat Island, Manning Reef, and Seal Rocks North. These loggers are exchanged annually and provide an ongoing record of *in-situ* water temperature and begin the process of developing an accurate climatology for the coral communities in the Harbour. The

available times series from deployed loggers were aggregated to weekly mean temperatures for presentation in this report. Recorded temperature is used to confirm the likelihood of thermal stress to corals in the Harbour estimated as degree heating weeks ([DHW tutorial](#)) published by the National Oceanic and Atmospheric Association [Coral Reef Watch](#). Maps of estimated maximum annual DHW for the past five years are provided to help visualise the relative thermal stress in recent years. Thresholds for likely bleaching are between 4-8 DHWs.

Degree Heating Weeks consider accumulated temperature anomalies that are greater than 1 degree C above the mean of the hottest monthly temperature for a location (Liu *et al.* 2017). The first three years of temperature data recorded by *in-situ* loggers has been used to estimate monthly mean temperatures over the 2017-2019 summers. The means of the hottest month from each year were used to provide a preliminary estimate of mean monthly maximums for each reef against which observed temperature profiles can be considered. Unfortunately, the logger deployed at Rat Island over the 2020-2021 summer was not recovered during surveys in 2021 as the picket to which it was attached had been dislodged and the logger lost.

3.4.2 Runoff

Runoff can impact corals in two ways: by introducing harmful loads of sediment and other contaminants and in extreme cases reducing salinity to levels lethal to corals (van Woesik 1991, Jones & Berkelmans 2014, Thompson *et al.* 2016). As a generalisation, the presence of coral communities can be interpreted as direct evidence that ‘typical’ salinity levels do not pose a threat to coral communities; it is deviations to levels below 28 parts per thousand (ppt) that begin to cause coral mortality (Berkelmans *et al.* 2012). As a first step in assessing the likelihood that floods exposed corals to low salinity or high loads of sediments and nutrients for the adjacent catchments the seasonal discharge of local rivers is compared to long term median flows. Median discharge for the “wet season”, defined here as December-May, is calculated from available data 1990-2010 and compared to the current year. Discharge data were sourced from the Queensland Government [water monitoring portal](#) for:

- Station 130005A-Fitzroy River at the Gap
- Station 132001A-Calliope River at Castlehope

As the flow of the Boyne River is interrupted by Lake Awoonga Dam, the time and magnitude of overflow of this dam, as reported by the [Gladstone Area Water Board](#), is also considered.

3.5 Plotting of results

3.5.1 Time-series of indicator and sub-indicators

The coral indicator scores over time for the whole-of-harbour and for each reporting zone are plotted as a time series based on data estimates returned to AIMS from the Data Integration and Management System (Figure 2). These plots include the mean and upper and lower confidence intervals based on bootstrapped distributions of the indicator scores. The observed values of the sub-indicators: coral cover, macroalgae and juvenile density (from which scores are derived) are plotted using their predicted trend and credible confidence limits estimated from generalised linear models that include random intercepts for each reef sampled (Figure 3, Figure 5, Figure 6).

3.5.2 *Comparison of Gladstone Harbour Coral communities to other near-shore reefs*

To place the state of the Gladstone Harbour coral communities into broader context, an ordination biplot has been provided (Figure 11). The biplot is based on Bray Curtis dissimilarity of square root transformed genus-level cover of hard and soft corals and higher-level groupings of algae observed on the Gladstone Harbour reefs in 2025, and at reefs sampled by the Marine Monitoring Program (MMP) in 2024. The MMP samples reefs using the same basic sampling methods as applied to the Gladstone Reefs. The MMP data used were the most recent observations available from 2 m depth sites.

To further compare Gladstone Harbour sub-indicator values to those observed on other near-shore reefs, boxplots are provided which show the distribution values for the Coral cover, Juvenile density and Macroalgae cover sub-indicators in Gladstone Harbour and regions monitored by the MMP (Figure 12). For the Coral change sub-indicator scores are plotted. As with the ordination described above, the MMP data uses only the most recent data available from 2 m depth sites.

4 RESULTS

The Harbour-wide coral indicator score for 2025 is 0.09 (very poor, grade E) and has been on a downward trajectory since a peak score of 0.28 (grade D) observed in 2017 (Table 1, Figure 2). The decline in the harbour-wide coral indicator score since 2017 reflects declines in both the Mid Harbour and Outer Harbour reporting zones. Coral cover across the Harbour remains at very low levels, resulting in the continued ‘very poor’, assessment for this sub-indicator (Table 1, Table A 2, Figure 3). Macroalgae cover has remained at high levels and assessment for this sub-indicator remains ‘very poor’ even though Macroalgae cover declined at the outer harbour sites (Table 1, Table A 2, Figure 4, Figure 5). The density of juvenile hard corals has continued to decline and associated sub-indicator score remains ‘very poor’ (Table 1, Table A 3, Figure 6, Figure 7). The coral change sub-indicator score decreased to ‘very poor’ following a steep decline in the Outer Harbour where the score for 2025 for this indicator was 0 (Table 1, Table 4, Figure 8).

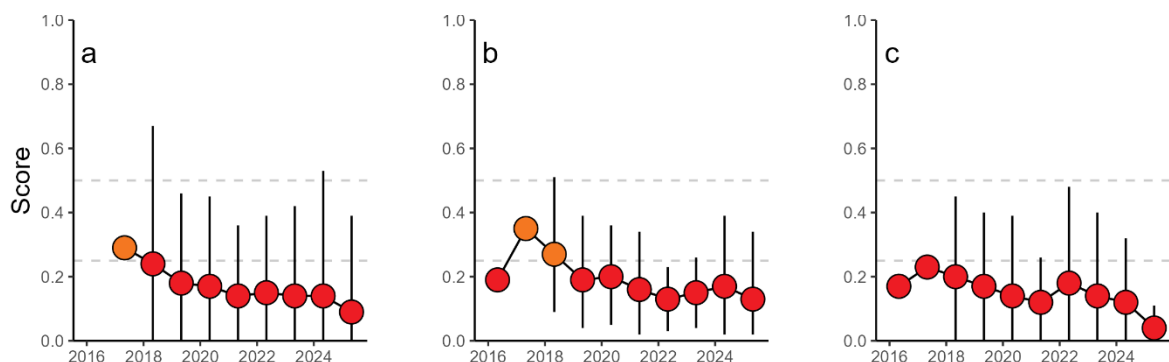


Figure 2 Coral indicator for reporting zones. a) Gladstone Harbour, b) Mid Harbour, c) Outer Harbour. Colours represent the coral report card grade (see Table 3 for details). Dashed lines indicate the thresholds between very poor, poor and satisfactory report card grades.

4.1 Coral cover

In 2025, mean coral cover decreased at all reefs except for at Rat Island where coral cover remained stable (Figure 3; Table A 6). However, the minor fluctuations observed over the six years of monitoring have all remained well within the levels categorised as ‘very poor’ for all reefs and the trend in this metric remains relatively flat across the Harbour (Figure 3a-c).

These results should be considered in terms of the threshold of 40% cover at which this indicator is categorised as ‘satisfactory’. This threshold approximates the baseline condition of a mean cover of 39% (maximum 47%) observed at reefs in the North Passage and along the western side of Facing Island in 2009 (BMT WBM 2013).

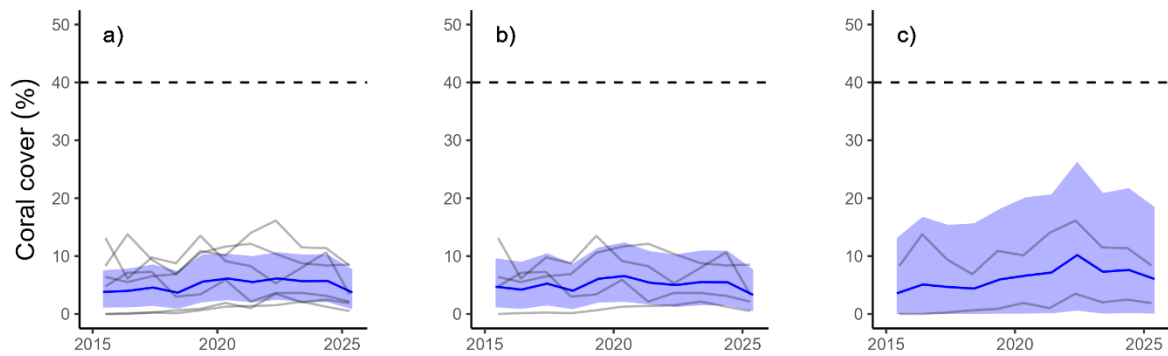


Figure 3 Trends in coral cover for reporting zones. a) Gladstone Harbour, b) Mid Harbour, c) Outer Harbour. Trends shown by blue lines bound by 95% confidence intervals of those trends (shading); grey lines represent observed profiles for individual reefs. Dashed lines represent the level at which this sub-indicator transitions between ‘poor’ and ‘satisfactory’ report card scores.

Extreme flooding of the Boyne River in 2013 caused Lake Awoonga to overflow and, in combination with flows from the Calliope River, will almost certainly have resulted in mortality of corals within the Harbour (Thompson *et al.* 2015, Jones *et al.* 2015). Monitoring of salinity within the Mid Harbour reporting zone by Vision Environment (2013a & b) confirmed modelling results (Jones *et al.* 2015) indicating the presence of water with salinity levels well below the threshold of 22 PSU, lethal to *Acropora* corals (Berkelmans *et al.* 2004), for a period of 3 days.

It is important not to over-interpret the minor inter-annual variability in coral cover observed since 2015. All coral estimates incur some degree of sampling error. The use of fixed transects does minimise this error, however some variability in estimates should be expected. Of note is that large erect species of macroalgae, can overtop corals excluding them from observation. High and variable cover of macroalgae is likely to have contributed to small inter-annual variations in coral cover but also slight underestimation of coral cover in general.

Within the Mid Harbour coral cover remains very low (Figure 3b). Rat Island’s trend of increase from 2016 to 2021 was interrupted with coral cover declining from 2021 to 2023 and stabilising in 2024 and 2025. There was a concomitant increase in macroalgae which has persisted through to 2025 (Figure 4). At both Facing Island and Farmers Reef coral cover has been variable with no trends evident. Corals remain very rare at Manning Reef where cover is limited to the few corals that have recruited in recent years (Figure 4).

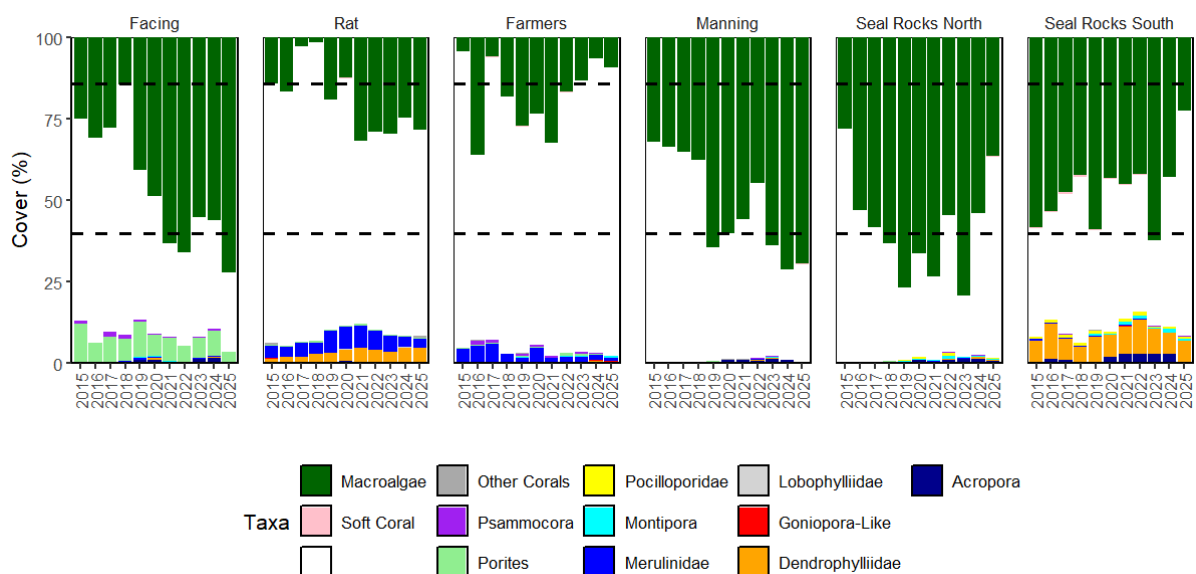


Figure 4 Composition of benthic cover at each location. Rising bars break down coral cover into major taxonomic groups (Families and Genera). Hanging bars represent macroalgae cover and are read in reverse (observed cover is read as 100% – y axis value, i.e. 10% cover will appear as a bar between 100% and 90% on the plot). White space is the remaining cover not occupied by indicators and will include: sand and silt substrate, turfing and crustose coralline algae along with other organisms such as sponges. Dashed reference lines indicate the boundary between the condition categories ‘Poor’ and ‘Satisfactory’. Hanging macroalgae cover bars not extending to the upper dashed reference line would be categorised as ‘Satisfactory’, or better. Rising bars for coral cover would have to extend to the lower dashed reference line to receive a ‘Satisfactory’ categorisation.

In the Outer Harbour, coral cover at Seal Rocks North remains extremely low amongst very high cover of macroalgae (Figure 4). At Seal Rocks South coral cover had steadily increased between 2018 and 2022, a continued decline has been evident since. The appearance of the genus *Acropora* within the coral community at Seal Rocks in 2020 (Figure 4) was an important step in the recovery of these communities as they have the potential to grow rapidly, and prior to floods in 2013 *Acropora* were a key component of the coral community (BMT WBM 2013). However, the cover of *Acropora* remained stable at low levels from 2020 until they disappeared again in 2025, demonstrating the ongoing environmental stress being imposed on these corals (Figure 4).

Scuba search data indicates that the bio-eroding sponge *Cliona orientalis* continues to impact the coral community across the Harbour and in particular colonies of *Turbinaria* at Seal Rocks South, along with *Porites*, *Favites*, *Psammocora*, *Plesiastrea* and *Cyphastrea* on Mid Harbour reefs (Table A 11). This sponge is almost certainly contributing to a lack of coral cover recovery across the Harbour. The presence of this sponge was first recorded at Manning Reef in 2024, due mostly to the previous absence of live coral, and persists in 2025 (Table A 11).

4.2 Macroalgae cover

The mean cover of macroalgae across the harbour continues to be very high (Figure 5a-c) ensuring the assessment for this sub-indicator remains ‘very poor’. Macroalgae cover increased slightly at Facing Island, Farmers Reef and Rat Island, and decreased at Manning Reef, Seal Rocks North and Seal Rocks

South (Figure 4). Despite the increase in macroalgae, Farmers Reef is the only site across the harbour to receive a 'good' categorisation for this indicator while all other reefs scored 'very poor', with Manning Reef and both sites at Seal Rocks consistently scoring 0 across the time series (Table A 6, Table A 7).

The generally high cover across the Harbour suggests that despite water quality being generally within guideline values in both the Mid and Outer Harbour ([2024 | Gladstone Healthy Harbour Partnership \(ghhp.org.au\)](https://ghhp.org.au)), the availability of nutrients within the Harbour is clearly not limiting to the macroalgae communities. Given the persistent high cover of macroalgae, and the performance of the other indicators, it is very likely that these algal communities are contributing to the suppression of coral recovery across the Harbour.

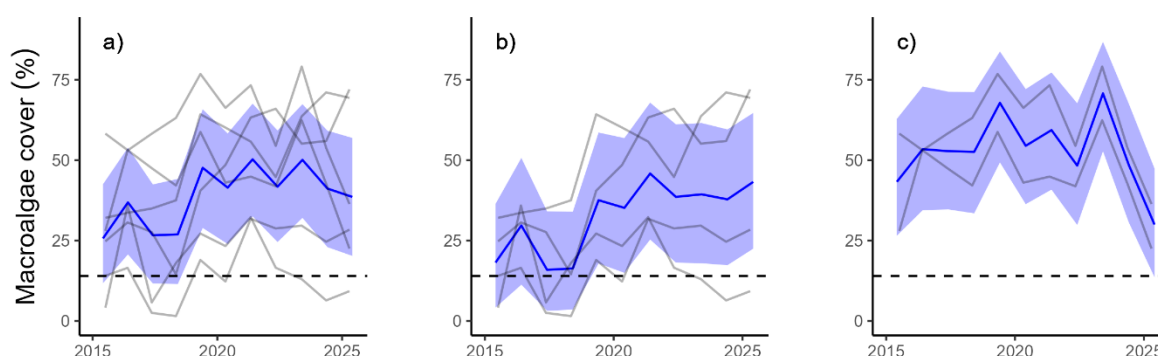


Figure 5 Trends in macroalgae cover for reporting zones. a) Gladstone Harbour, b) Mid Harbour, c) Outer Harbour. Trends shown by blue lines bound by 95% confidence intervals of those trends (shading); grey lines represent observed profiles for individual reefs. Dashed lines represent the level at which this sub-indicator transitions between 'poor' and 'satisfactory' report card scores.

As with coral communities (Table A 9) differences in the taxonomic composition of macroalgal communities (Table A 10) suggest fine scale differences in the combined physical and chemical environments at the monitoring locations. Monitoring undertaken by the MMP elsewhere on the GBR demonstrates that at reefs predisposed to high levels of macroalgae, cover is typically variable between years (Thompson *et al.* 2025). Within Gladstone Harbour, variability in macroalgae communities is especially evident at reefs in the Mid Harbour zone where cover and composition vary both from year to year within individual reefs, and between reefs (Figure 5b, Table A 10). In contrast, although there is some variability in the overall cover of macroalgae, the community composition at reefs in the Outer Harbour appear relatively stable, with communities consistently dominated by the brown macroalgae family Sargassaceae, and the genus *Lobophora* (Table A 10).

4.3 Juvenile density

The harbour-wide mean density of juvenile corals remains very low and well below levels observed in early years of the program ensuring that this sub-indicator remains 'very poor' in 2025 (Figure 6a, Table A 3). Notably lacking across most Mid Harbour reefs are juveniles of the family Acroporidae (Figure 7). High coral cover recorded at several sites in 2009 included a high representation of this family (BMT WBM 2013). Indeed, dead *Acropora* skeletons are the primary substrate at both Seal Rocks North and Manning Reef (authors pers. obs.). Until juveniles of this family appear and survive, recovery of coral communities at these locations is unlikely.

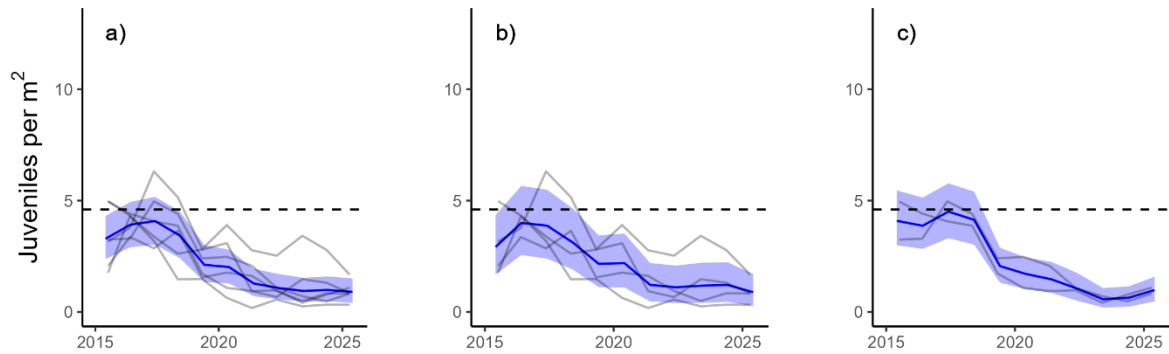


Figure 6 Trends in juvenile density for reporting zones. a) Gladstone Harbour, b) Mid Harbour, c) Outer Harbour. Trends shown by blue lines bound by 95% confidence intervals of those trends (shading); grey lines represent observed profiles for individual reefs. Dashed lines represent the level at which this sub-indicator transitions between ‘poor’ and ‘satisfactory’ report card scores.

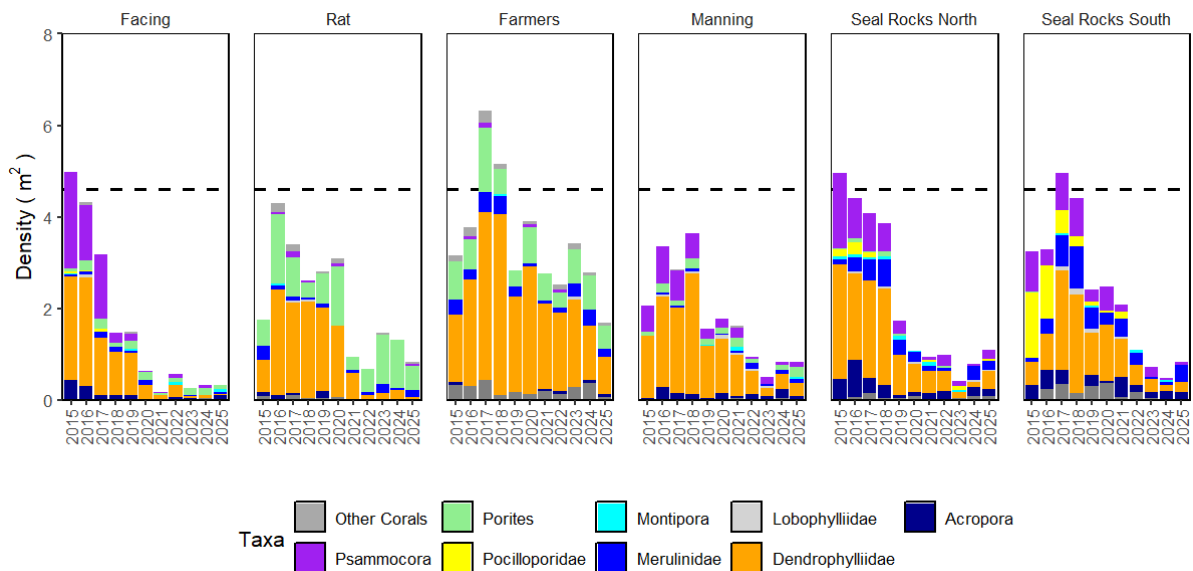


Figure 7 Composition of juvenile coral communities at each location. Bars break down juvenile density into major taxonomic groups (Families and Genera). Dashed reference line indicates the boundary between the condition categories ‘Poor’ and ‘Satisfactory’. Juvenile density would have to extend to the reference line to receive a ‘Satisfactory’ categorisation.

4.4 Change in hard coral cover

The Harbour-level score for coral cover change has declined from ‘poor’ to ‘very poor’ for the first time since this indicator was first estimated in 2017 (Figure 8a). The Mid Harbour score has declined since 2021 to the ‘very poor’ category which continues in 2025 (Figure 8b). ‘Very poor’ scores returned for Farmers Reef, Manning Reef and Rat Island are most influential in this result (Table 4). In the Outer Harbour the score decreased to ‘very poor’ for the first time since surveys began (Figure 8c). This was driven by Seal North decreasing significantly from a ‘satisfactory’ score of 0.5 in 2024 to a ‘very poor’ score of 0 in 2025, and Seal Rocks South declined further from 0.14 in 2024 to 0 in 2025 (Table 4, Table

A 7). It should be noted that the ‘satisfactory’ score for Seal Rocks North in 2024 was influenced by the very limited expectation of change for a reef with such low coral cover.

The Harbour wide decline in coral cover change score for 2025 indicates that these corals are undergoing significant environmental stress. This score is calculated using a four-year rolling mean, so a positive score from four years ago will have now moved out of the calculation, causing a sharper drop in the score for this year. In previous years, the ongoing Harbour-wide ‘poor’ coral cover change score demonstrates that recovery of coral communities continues to fall short of modelled expectations (Figure 8a). Important to note is that the scores for this indicator are based on observations of coral cover recovery averaged over a three-year period. Explicitly excluded are inter-annual changes in hard coral cover deemed to have been influenced by an acute pressure. For example, the cover change score for the Outer Harbour in 2025 is based only changes in hard coral cover observed between 2022 and 2023, as subsequent changes were deemed to have been influenced by the immediate and longer-term impacts of high water temperatures that caused coral bleaching in 2024.

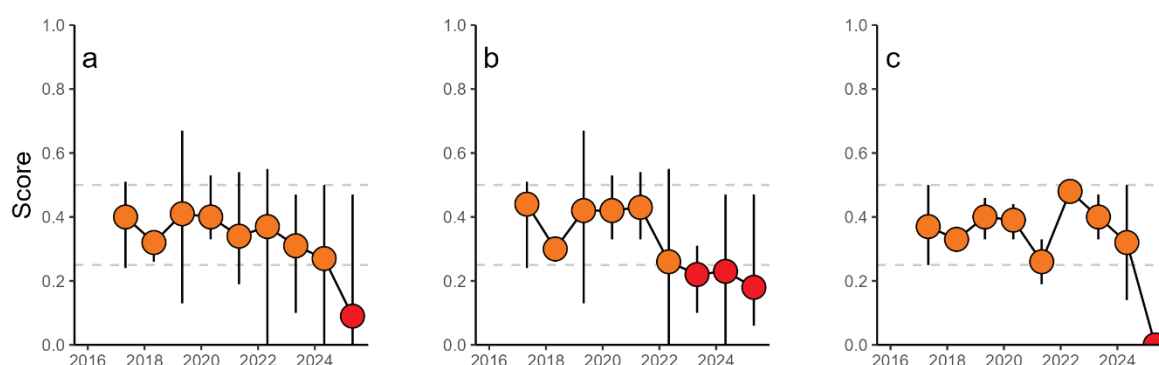


Figure 8 Coral cover change timeseries. Scores for a) Harbour, b) Mid Harbour and c) Outer harbour. Dashed reference lines included for lower thresholds for ‘Satisfactory’ and ‘Poor’ condition categorisations.

Table 4 Coral cover change sub-indicator scores for reefs and zones 2025.

Zone	Reef	Reef-level		Zone-level	
		Score	Condition	Score	Condition
Mid Harbour	Facing Island	0.47	Poor	0.18	Very Poor
	Farmers Reef	0.09	Very Poor		
	Manning Reef	0.10	Very Poor		
	Rat Island	0.06	Very Poor		
Outer Harbour	Seal Rocks North	0	Very Poor	0	Very Poor
	Seal Rocks South	0	Very Poor		

4.5 Environmental pressures

Seawater temperatures over the 2024/2025 summer were higher enough to have caused thermal stress to corals within the Harbour. *In situ* records show summer water temperatures exceeded the

short-term mean (2017-2019) but were slightly lower than temperatures observed in 2020 and 2024 when bleaching impacted corals across the harbour (Figure 9). Degree Heating Weeks (DHW, Liu *et al.* 2018) provide a summary of possible temperature related stress to corals. DHW estimates accumulate time of exposure of more than 1 degree C above the mean of the hottest month from a location's climatology. Where positive temperature anomalies occur in the hottest summer months DHW values will predict levels of bleaching probability. Degree Heating Week estimates for 2024/25 over the summer period (December to March inclusive) adjacent to Gladstone Harbour were 4 – 6 DHW, a level associated with possible coral bleaching (Figure 10). Although temperatures had decreased by the time of survey in April 2025, a small number of colonies at Manning and Seal Rocks South were still bleached (Table A 11).

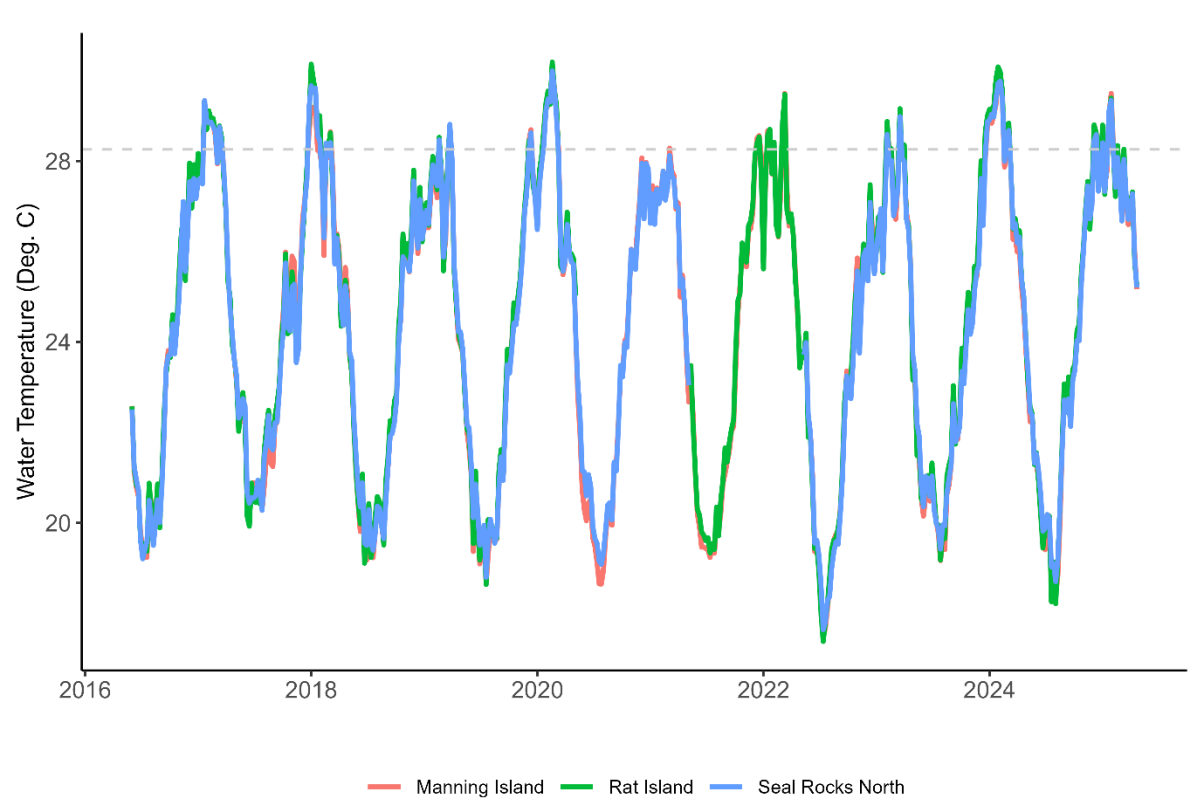


Figure 9. Weekly mean temperature profiles recorded at coral monitoring locations. Grey dashed horizontal reference line is included as a visual aid and represent the mean of the hottest months in 2017, 2018 and 2019 at each reef. The temperature logger for the 2021-2022 year at Seal Rocks North was lost.

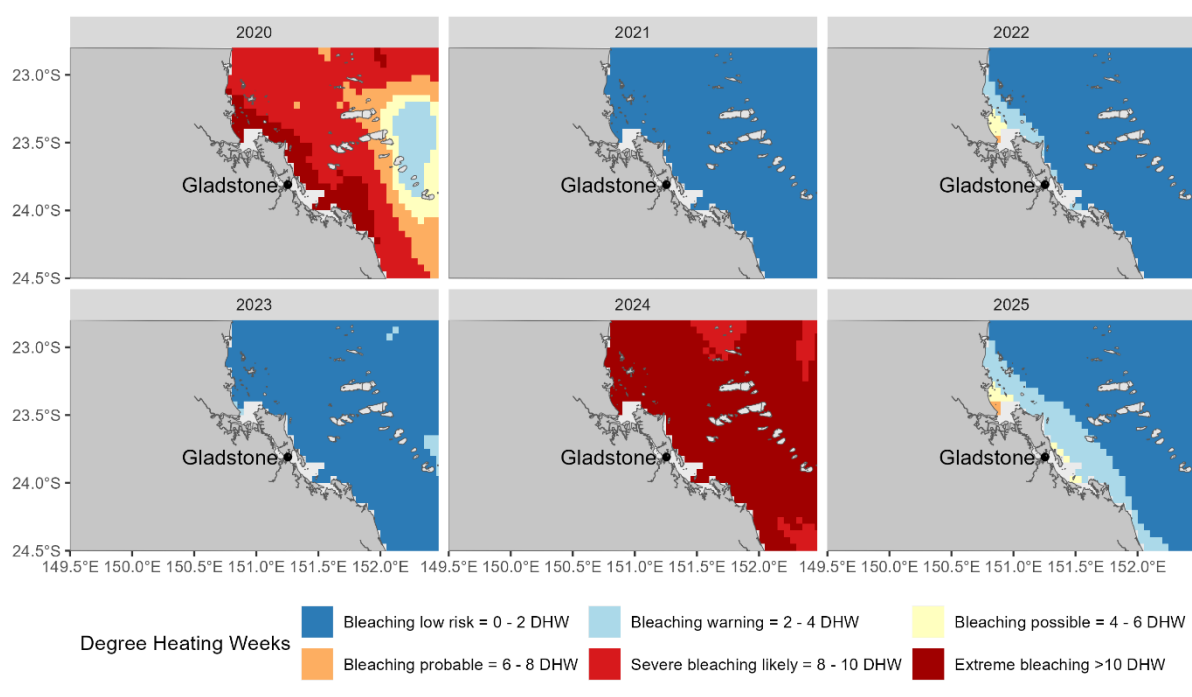


Figure 10 Annual maximum Degree Heating Week estimates.

River discharge for the 2024/2025 wet season was 1.3 and 2.2 times above the baseline median from the Calliope River and Fitzroy River respectively (Table 5). The Awoonga Dam levels have remained below the spillway since January 2018. Higher discharge from the Calliope and Fitzroy rivers suggests a potential for low salinity flood impacts and delivery of nutrients and sediments from the local catchments during the year preceding coral community surveys in 2025. However, the effect of this wet season discharge is expected to be significantly less than that following the extreme flooding of the Calliope River (Table 5) and overflow of the Lake Awoonga Dam in 2013 (as reported in Thompson et al. 2015). No evidence of mortality associated with low salinity was observed during surveys in 2025.

Table 5 River discharge. Values are annual wet season (December to May) discharge as a multiple of the baseline median wet season discharge for the period (1990-2010).

River	Median (ML)	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025
Calliope	53306	17	2.8	5.2	1.2	4.4	0.5	0.0	0.9	0.2	0.7	0.3	0.3	1.3
Fitzroy	1540100	5.5	1.0	1.7	1.5	3.9	0.5	0.9	1.6	0.3	1.7	1.3	1.2	2.2

4.6 Comparisons with other near-shore reefs

The composition of benthic communities within Gladstone Harbour are distinct from communities on most near-shore reefs monitored by the MMP. Ordination analysis highlights it is the combination of low cover of most coral genera and high cover of macroalgae that define the Gladstone Harbour communities (Figure 11). The primary axis of the ordination (MDS1, Figure 11) explains 36% of the

variance in community composition and effectively separates reefs with communities dominated by red or brown macroalgae on the right from those with proportionally higher cover of a range of coral genera, to the left. The distinction evident in the ordination is clear when comparing the relatively low coral cover and high macroalgae cover on Gladstone Harbour reefs to the cover of these groups at near-shore reefs in other regions (Figure 12a, c).

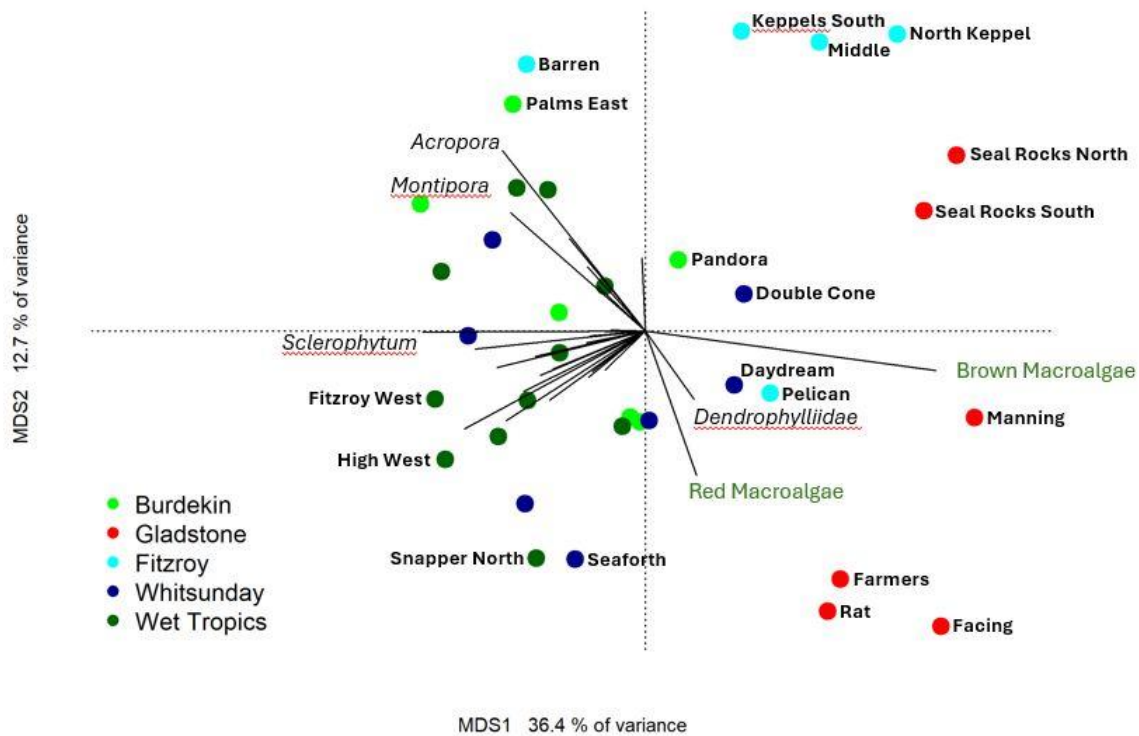


Figure 11 Ordination biplot comparing Gladstone Harbour reef communities with other near-shore reefs. The biplot is based on Bray-Curtis dissimilarity of square root transformed cover of coral (genus or family – vectors with black text italicised) and higher order-level cover of macroalgae (vectors with green text) observed on the Gladstone Harbour reefs in 2025 and at 2 m reefs sampled by the Marine Monitoring Program (MMP) in 2024. Labels on the axis indicate how much of the variance in community composition is captured by the first (x-axis, MDS1) and second (y-axis, MDS2) dimensions of the ordination. Reef names are black bolded text, and the reefs are coloured by their NRM Region. Note that Snapper South in the Wet Tropics has been removed due to disturbance reducing hard coral cover and most macroalgae to 0 during the 2024 wet season.

As might be expected, the Gladstone Harbour reef community composition mostly resembles those in the Fitzroy Region (Figure 11). Most reefs monitored in the Fitzroy Region were grouped closely with those in Gladstone Harbour due to comparatively high cover of macroalgae relative to most corals. The one coral group that remains common on most Fitzroy Region reefs is *Acropora*, leading to the distinction between Fitzroy Region and Gladstone Harbour coral communities on the Y- axis.

Like the reefs in Gladstone Harbour, recovery of coral communities in the Fitzroy region has been low and macroalgae continue to dominate the benthic community, especially as the Fitzroy region was severely impacted by the 2024 summer bleaching event (Thompson *et al.* 2025). The benthic communities at two reefs in the Whitsunday Region, [Daydream Island](#) and [Double Cone Island](#), also resemble Gladstone reefs. Both these reefs were severely impacted by Cyclone Debbie in 2017, with

coral being reduced to very low levels. As with the Gladstone Harbour reefs, the benthic communities at Daydream Island and Double Cone Island underwent a distinct increase in the cover of macroalgae and had shown negligible recovery in 2024 at Double Cone, while juvenile coral density is increasing every year at Daydream (Thompson *et al.* 2025).

The reefs of Gladstone Harbour sit in the lower end of the spectrum for most indicators relative to other near-shore reefs monitored by the MMP (Figure 12). Whilst it is evident that these have supported healthier coral communities in the past (BMT WBM 2013), present coral cover is well below that of other inshore areas.

Of concern is the continued low density of juvenile corals and high cover of macroalgae within Gladstone Harbour compared to other inshore reefs, which present a bottleneck to recovery (Figure 12 b, c).

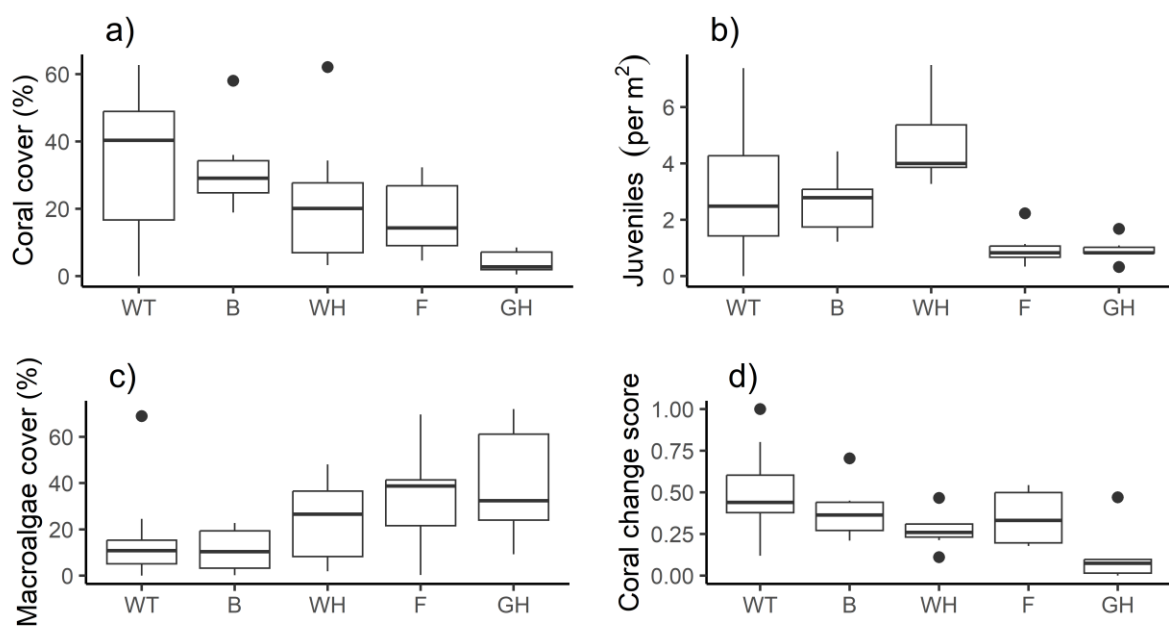


Figure 12 Regional comparisons of sub-indicators. a) coral cover, b) juvenile density, c) macroalgae cover, d) coral change score. WT= Wet Tropics, B=Burdekin, WH=Whitsundays, F= Fitzroy, GH= Gladstone Harbour. Data are based on current indicator values and scores for Gladstone Harbour reefs and the most recent available data for 2 m reefs sampled by the MMP.

The mean coral change indicator is much lower than has been reported in any other region by the MMP in 2024, but is most similar to the Whitsunday region which is still recovering slowly from the impacts of cyclone Debbie (Figure 12d). In the Whitsundays, several reefs had shown little to no recovery of coral cover since being severely impacted by cyclone Debbie, however juvenile coral density, especially at Daydream Island is increasing (Thompson *et al.* 2024). In the Gladstone region, juvenile corals remain sparse, limiting recovery of these reefs (Figure 12). Further, at most shallow reef sites in the Whitsunday Region, the coral communities include higher representation of relatively fast-growing corals, and this influences the expected rate of coral cover at those reefs. Finally, the MMP scoring system is biased toward lower scores compared to that used for the Gladstone scores as changes in cover equivalent to the lower confidence interval of predicted change returns a score of 0.4 within the MMP system, compared to 0.5 for Gladstone. In combination, this means that while

scores were closest to those observed in the Whitsunday Region, the observed rate of recovery remains low within the Harbour.

5 CONCLUSION

Results from surveys in 2025 demonstrate a further decline in the condition of coral communities in Gladstone Harbour. While coral bleaching, in response to high water temperature in early 2020 and again in 2024, is likely to have contributed to the current very poor condition, ongoing monitoring since 2015 demonstrates a clear lack of recovery since the severe loss of coral noted in 2015 (Thompson *et al.* 2015). This loss of coral coincided with the inundation of the Harbour's reefs by floodwaters in 2013. The low salinity within the harbour during these floods was considered the primary cause of coral mortality and precluded the ability to consider any cumulative or prior impact of extensive development of the Harbour through to 2013 (Jones *et al.* 2015, Thompson *et al.* 2015).

The magnitude of coral loss that occurred in 2013 dictated very low scores for the coral cover sub-indicator in 2015. Following such severe disturbance, it is the recovery processes that best describe the coral communities' overall condition. The Macroalgae cover, Juvenile density, and Coral change sub-indicators are all formulated to assess the recovery process and, collectively, demonstrate the limited recovery potential continuing to be exhibited by the coral communities within the Harbour.

In combination, the continued 'very poor' scores for each sub-indicator corroborate studies that demonstrate density-dependant feedback mechanisms which promote a persistent shift from coral to macroalgal dominance where conditions allow the proliferation of macroalgae (Mumby *et al.* 2007, Mumby *et al.* 2013). Across the inshore reefs monitored by AIMS, the prevalence of macroalgae in shallow waters was positively related to Chlorophyll *a* and total suspended solids concentration in the surrounding waters, implicating poor water quality as a factor contributing to their prevalence (Thompson *et al.* 2025). Macroalgae are abundant across the harbour, and large fleshy macroalgae such as family Sargassaceae, the genus *Asparagopsis* and, the low mat-forming genera *Lobophora* and *Dictyota* have been shown to be highly disruptive to coral community recovery (Birrell *et al.* 2008, Diaz-Pulido *et al.* 2010, Hauri *et al.* 2010). Macroalgae such as *Lobophora* and *Dictyota* have been shown to directly impact adult coral colonies leading to tissue loss, declines in coral fitness, and reduced growth rate (Lirman 2001, Vega Thurber *et al.* 2012, Morrow *et al.* 2017). Ongoing competition between coral and macroalgae is likely to contribute to the 'very poor' score for the coral change sub-indicator at most reefs. High cover of macroalgae is also likely to be affecting coral recruitment processes contributing to the 'very poor' score for juvenile density (Johns *et al.* 2018, Page *et al.* 2023, Burgo *et al.* 2025). High cover of macroalgae has been associated with low densities of juvenile corals on several reefs monitored by the MMP. Notable examples include some reefs in the Fitzroy Region where the density of juvenile corals has remained very low since high cover of macroalgae replaced corals killed by floodwaters in 2011 (Berkelmans *et al.* 2012, Thompson *et al.* 2025).

In addition to hampering recruitment, coral-macroalgae interactions can potentially reduce the fecundity of adult corals (Tanner 1995, Foster *et al.* 2008), and this may be further limiting the supply of larvae from an already depleted population of adult corals.

Ongoing low density of juvenile corals indicates that recruitment processes and/or post-recruitment survivorship are a severe bottleneck for the recovery of these coral communities. Generally, low recruitment of corals may be expected given the low abundance of coral brood-stock within the Harbour that will naturally limit local population fecundity. The Allee Effect - low rates of fertilization

due to the low density of spawning corals, will further reduce the supply of locally spawned larvae (Gascoigne & Lupcius 2004). Coral recruitment can also be suppressed by a range of water quality constituents, including fine-grained sediments, metals and chemical contaminants (Richmond *et al.* 2018). It is beyond the scope of the available data to tease apart the cumulative influences of population-level and water quality related pressures on coral recruitment within the Harbour. That said, investigating avenues to reduce anthropogenic impacts during the annual period of coral spawning through to settlement would seem prudent (Fraser *et al.* 2017).

The decline of juvenile coral densities in recent years is likely influenced by multiple processes. In addition to the ongoing influence of macroalgae, as described above, high water temperatures in early 2020 and 2024 are likely to have reduced juvenile densities by increasing the mortality rate of settled corals or potentially limiting the fecundity of adult corals over the later end of the spawning season (Ward *et al.* 2002). An examination of the composition of juvenile communities highlights that it is the genus *Turbinaria* that has undergone the greatest absolute reduction across the Harbour. High variability in the density of *Turbinaria* juveniles has also been noted at some turbid water reefs monitored elsewhere on the Great Barrier Reef by the MMP (Thompson *et al.* 2025), although the underlying process driving this variability remains unknown. The juvenile communities across the Harbour do, however, continue to include a higher diversity of genera than the adult communities, suggesting connectivity of larvae from beyond the Harbour. Although these juveniles are yet to contribute to increased coral cover, this apparent connectivity to more distant brood-stock is a promising sign for the resilience of these communities. The continued presence of *Acropora* juveniles, although in low densities and fluctuating from year to year, remains a positive sign. *Acropora* were a key component of the coral communities at most sites prior to the 2013 floods (BMT WBM 2013), and the reestablishment of these fast-growing species will be fundamental to the recovery of these communities.

The coral change sub-indicator explicitly accounts for an expected low rate of coral cover increase due to both the low existing coral cover and communities dominated by slow growing species. Despite these modest expectations, the poor scores for this metric demonstrate that combined pressures imposed by the environmental conditions within the Harbour are limiting the recovery of coral cover. Further influencing the score for this sub-indicator is the widespread presence of the bio-eroding sponge *Cliona orientalis* which continues to be the most significant contributor to coral mortality within the Harbour.

High water temperatures in early 2020 and 2024 were a further setback to the recovery of coral communities. The stress caused by high temperatures was particularly evident at the Outer Harbour sites where surveys in late April 2020 revealed that many colonies of the families Acroporidae and Pocilloporidae were at least partially bleached (Costello *et al.* 2020). These families are comprised of relatively fast-growing species key to the recovery of coral cover. Lower influence of bleaching in the Mid Harbour is likely due to the higher proportion of bleaching-resistant taxa at those sites. In the summer of 2023/24, the region saw high water temperatures resulting in over 10 degree-heating weeks which is associated with extreme coral bleaching. Bleaching of individual colonies was still occurring in the Harbour during surveys conducted in May, similar to reports from One Tree Reef (situated around 43nm east-northeast from Gladstone) where 31% of corals were still bleached in July 2024 (Byrne *et al.* 2025). While the temperatures were not as extreme over the 2024/25 summer season, when surveys were conducted in April of 2025 individual colonies were still bleached, indicating on-going thermal stress.

In the broader context of near-shore reefs on the GBR, comparison of sub-indicator scores with those from reefs in other regions demonstrate Gladstone Harbour reefs perform poorly. In general, the Gladstone Harbour reefs were most like reefs which had recently undergone severe disturbances or had not recovered from previous disturbances (Figure 11). These communities exhibit either very low coral cover, very high cover of macroalgae or a combination of both. Of concern is that at Pelican Island in the Fitzroy Region, where benthic communities are most like the Gladstone reefs, the coral community has shown negligible recovery since 2011 (Thompson *et al.* 2025). What is unclear is whether such long-term hiatus in recovery is natural for coral communities in these marginal conditions or symptomatic of delayed recovery in the face of mounting cumulative pressures associated with climate change and declining water quality.

Overall, the 2025 results further demonstrate the continued lack of resilience in coral communities within Gladstone Harbour, which has been previously reported (Thompson *et al.* 2024). Given the current depleted state of coral cover, recovery will depend heavily on the connectivity with reefs beyond the Harbour for larval supply. However, the subsequent settlement and growth of these larvae are likely to remain low until the underlying conditions promoting the continued high cover of macroalgae are identified and mitigated.

The clear lack of coral community recovery over a decade of monitoring suggests that reefs within the harbour are in a state of hysteresis whereby the high cover of macroalgae relative to corals is being perpetuated. High cover of macroalgae on the GBR is generally limited to the relatively turbid and nutrient rich inshore areas (De'ath & Fabricius 2010). However, within this habitat shifts from coral to macroalgae dominated communities are often caused by an acute event that kills corals, providing space that is rapidly colonised by macroalgae (Ceccarelli *et al.* 2020). Once established, the prevalence of macroalgae is supported by density dependant feedback processes that limit recruitment, growth, and survival of hard corals. Central to returning to a coral dominated state is increasing the supply of new coral colonies into the system (Hock *et al.* 2025). While increased supply may occur naturally, due to variable supply and or survival of coral larvae, targeted management interventions are likely to increase the likelihood of the system returning to a coral dominated state (Hock *et al.* 2025). Given the very low populations of coral and high cover of macroalgae across Gladstone Harbour, the recovery of coral communities may require a multi-pronged management plan that includes both providing a source of larval or juvenile corals and removal of macroalgae to enhance their survival. The active removal of macroalgae species common to Gladstone Harbour, and in particular *Sargassum* has been shown to increase both the settlement of coral larvae and growth of adult corals elsewhere in the inshore GBR (Smith *et al.* 2023, Burgo *et al.* 2025) proffering a potential avenue for coral reef restoration within the harbour. At the same time, as the distribution of macroalgae within the Great Barrier Reef is broadly governed by water quality, efforts to improve water quality may also prove beneficial.

Finally, the restoration of coral communities within the inshore GBR is in its infancy and it would be advantageous to design any assessment of the efficacy of any interventions to leverage the information within the existing coral monitoring program.

6 ACKNOWLEDGMENTS

This report continues a series of annual reports. We acknowledge Paul Costello as previous author and Stephen Neale who was integral to the collection of field data in 2021. We note also the valuable insight provided by Mike Emslie and the independent science panel who's comments on previous drafts improved this report.

7 REFERENCES

- Berkelmans R, De'ath G, Kininmonth S, Skirving WJ (2004) A comparison of the 1998 and 2002 coral bleaching events on the Great Barrier Reef: spatial correlation, patterns and predictions. *Coral Reefs* 23:74-83. doi: 10.1007/s00338-003-0353-y
- Berkelmans R, Jones AM, Schaffelke B (2012) Salinity thresholds of *Acropora* spp. on the Great Barrier Reef. *Coral Reefs* 31:1103-1110. doi: 10.1007/s00338-012-0930-z
- Birrell CL, McCook LJ, Willis BL, Diaz-Pulido GA (2008) Effects of benthic algae on the replenishments of corals and the implications for the resilience of coral reefs. *Oceanography and Marine Biology: An Annual Review* 46:25-64
- BMT WBM (2013) Central Queensland corals and associated benthos: Monitoring review and gap analysis. April 2013. Prepared for the Gladstone Ports Corporation.
- Burgo M, Fabricius KE, Hoey AS (2025) The structure and composition of macroalgal communities influence coral recruitment on an inshore reef of the Great Barrier Reef. *Coral Reefs* 44:1315-1326. doi: 10.1007/s00338-025-02691-0
- Byrne M, Waller A, Clements M, Kelly AS, Kingsford MJ, Liu B, Reymond CE, Vila-Concejo A, Webb M, Whitton K, Foo SA (2025) Catastrophic bleaching in protected reefs of the Southern Great Barrier Reef. *Limnology and Oceanography Letters* 10:340-348. doi: 10.1002/lol2.10456
- Ceccarelli DM, Evans RD, Logan M, Mantel P, Poutinen M, Petus C, Russ GR, Williamson DH (2020) Long-term dynamics and drivers of coral and macroalgal cover on inshore reefs of the Great Barrier Reef Marine Park. *Ecological Applications* 30(1):e02008. doi: 10.1002/eap.2008
- Cheal AJ, Emslie M, MacNeil MA, Miller I, Sweatman H (2013) Spatial variation in the functional characteristics of herbivorous fish communities and the resilience of coral reefs. *Ecological Applications* 23(1):174-188. doi: 10.1890/11-2253.1
- Costello P, Thompson A, Davidson J (2020) Coral Indicators for the 2020 Gladstone Harbour Report Card 2020: ISP014. Report prepared for Gladstone Healthy Harbour Partnership. Australian Institute of Marine Science, Townsville. (46 pp)
- De'ath G, Fabricius K (2010) Water quality as a regional driver of coral biodiversity and macroalgae on the Great Barrier Reef. *Ecological Applications*. 20(3):840-850. doi: 10.1890/08-2023.1

- De'ath G, Fabricius KE, Sweatman H, Puotinen M (2012) The 27-year decline of coral cover on the Great Barrier Reef and its causes. *Proceedings of the National Academy of Sciences* 190(44):17995-17999
- Diaz-Pulido G, Harii S, McCook LJ, Hoegh-Guldberg O (2010) The impact of benthic algae on the settlement of a reef-building coral. *Coral Reefs* 29:203-208. doi: 10.1007/s00338-009-0573-x
- Erftemeijer PLA, Reigl B, Hoeksema BW, Todd PA (2012) Environmental impacts of dredging and other sediment disturbances on corals: A review. *Marine Pollution Bulletin* 64(9):1737-1765. doi: 10.1016/j.marpolbul.2012.05.008
- Fabricius KE (2005) Effects of terrestrial runoff on the ecology of corals and coral reefs: Review and synthesis. *Marine Pollution Bulletin* 50(2):125-146. doi: 10.1016/j.marpolbul.2004.11.028
- Fraser MW, Short J, Kendrick G, McLean D, Keesing J, Byrne M, Caley MJ, Clarke D, Davis AR, Erftemeijer PL, Field S, Gustin-Craig S, Huisman J, Keough M, Lavery PS, Masini R, McMahon K, Mengersen K, Rasheed M, Statton J, Stoddart J, Wu P (2017) Effects of dredging on critical ecological processes for marine invertebrates, seagrasses and macroalgae, and the potential for management with environmental windows using Western Australia as a case study. *Ecological Indicators*. 78:229-42. doi: 10.1016/j.ecolind.2017.03.026
- Foster NL, Box SJ, Mumby PJ (2008) Competitive effects of macroalgae on the fecundity of the reef-building coral *Montastrea annularis*. *Marine Ecology Progress Series* 367: 143-152. doi: 10.3354/meps07594
- Garde LA, Spillman CM, Heron S, Beeden R (2014) ReefTemp Next Generation: A new operational system for monitoring reef thermal stress. *Journal of Operational Oceanography* 7:21-33. doi: 10.1080/1755876X.2014.11020150
- Gascoigne J, Lipcius MR (2004) Allee effects in marine systems. *Marine Ecology Progress Series* 269: 49-59 doi: 10.3354/meps269049
- Hauri C, Fabricius K, Schaffelke B, Humphrey C (2010) Chemical and physical environmental conditions underneath mat- and canopy-forming macroalgae, and their effects on understory corals, *PLoS ONE* 5(9):e12685 doi: 10.1371/journal.pone.0012685
- Hock K, Hastings A, Doropoulos C, Babcock RC, Oritz JC, Thompson A, Mumby PJ (2024) Transient dynamics mask the resilience of coral reefs. *Theoretical Ecology* 17:1-12. doi: 10.1007/s12080-023-00570-4
- Hughes TP, Rodrigues MJ, Bellwood DR, Ceccarelli D, Hoegh-Guldberg O, McCook L, Moltschaniwskyj N, Pratchett Ms, Steneck RS, Willis B (2007) Phase shifts, herbivory, and the resilience of coral reefs to climate change. *Current Biology* 17(4):360-365. doi: 10.1016/j.cub.2006.12.049
- Johns KA, Emslie MJ, Hoey AS, Osborne K, Jonker MJ, Cheal AJ (2018) Macroalgae feedbacks and substrate properties maintain a coral reef regime shift. *Ecosphere* 9(7):e02349. doi: 10.1002/ecs2.2349
- Jones AM, Berkelmans R (2014) Flood impacts in Keppel Bay, Southern Great Barrier Reef in the aftermath of cyclonic rainfall. *PLoS ONE* 9(1): e84739. doi: 10.1371/journal.pone.0084739

- Jones CM, Richardson DL, Baheerathan R, Guard PA, Ettema S (2015) Prioritisation of reef restoration and enhancement sites – Phase 2 and 3 report. Report produced for Gladstone Ports Corporation's Biodiversity Offset Strategy. 130pp
- Jonker MJ, Bray PE, Johns KA, Osborne KO (2020) Surveys of benthic reef communities using underwater digital photography and counts of juvenile corals. Long Term Monitoring of the Great Barrier Reef - Standard Operational Procedure Number 10, 2nd ed. Australian Institute of Marine Science, Townsville, Australia. 36 pp. doi: 10.25845/jjzj-0v14
- Kroon FJ, Kuhnert PM, Henderson BL, Wilkinson SN, Kinsey-Henderson A, Abbott B, Brodie JE, Turner RDR (2012) River loads of suspended solids, nitrogen, phosphorus and herbicides delivered to the Great Barrier Reef lagoon. *Marine Pollution Bulletin* 65:167–181. doi: 10.1016/j.marpolbul.2011.10.018
- Lirman D (2001) Competition between macroalgae and corals: effects of herbivore exclusion and increased algal biomass on coral survivorship and growth. *Coral Reefs* 19:392-399. doi: 10.1007/s003380000125
- Liu G, Skirving WJ, Geiger EF, De La Cour JL, Marsh BL, Heron SF, Tirak KV, Strong AE, Eakin CM (2017) NOAA Coral Reef Watch's 5km Satellite Coral Bleaching Heat Stress Monitoring Product Suite Version 3 and Four-Month Outlook Version 4. *Reef Encounter* 45 32(1): 39-45
- Liu G, Strong AE, Skirving W, Arzayus LF (2006) June. Overview of NOAA coral reef watch program's near-real time satellite global coral bleaching monitoring activities. In *Proceedings of the 10th international coral reef symposium* (Vol. 1793:1783-1793)
- McCook LJ, Jompa J, Diaz-Pulido G (2014) Competition between corals and algae on coral reefs: a review of evidence and mechanisms. *Coral Reefs* 19:400-417. doi: 10.1007/s003380000129
- Miller IR, Jonker M, Coleman G (2009) Crown-of-thorns seastars and coral surveys using the manta tow and SCUBA Search techniques. Long-term Monitoring of the Great Barrier Reef Standard Operational Procedure Number 9, Edition 3. Australian Institute of Marine Science, Townsville
- Morrow KM, Bromhall K, Motti CA, Munn CB, Bourne DG (2017) Allelochemicals produced by brown macroalgae of the *Lobophora* genus are active against coral larvae and associated bacteria, supporting pathogenic shifts to *Vibrio* dominance. *Appl Environ Microbiol* 83:e02391-16. doi: 10.1128/AEM.02391-16.
- Mumby PJ, Hastings A, Edwards HJ (2007) Thresholds and the Resilience of Caribbean Coral Reefs. *Nature* 450 (7166): 98–101. doi:10.1038/nature06252.
- Mumby PJ, Steneck RS, Hastings A (2013). Evidence for and against the Existence of Alternate Attractors on Coral Reefs. *Oikos* 122. doi:10.1111/j.1 600-0706.2012.00262.x.
- Osborne K, Dolman AM, Burgess SC, Johns KA (2011) Disturbance and the dynamics of coral cover on the Great Barrier Reef (1995–2009). *PLoS ONE* 6:e17516. doi: 10.1371/journal.pone.0017516
- Page CA, Giuliano C, Meehan K, Fisher R, Motti CA, Negri AP, Randall CJ (2023) Varied effects of *Lobophora* chemistry on settlement of larvae from five coral genera. *Marine Ecology Progress Series* 717:17-35. doi: 10.3354/meps14380
- Queensland Government (2015) Report Cards, <http://www.reefplan.qld.gov.au/measuring-success/report-cards/>

- Richmond RH, Tisthammer KH and Spies NP (2018) The Effects of Anthropogenic Stressors on Reproduction and Recruitment of Corals and Reef Organisms. *Frontiers in Marine Science* 5:226. doi: 10.3389/fmars.2018.00226
- Schleussner CF, Lissner TK, Fischer EM, Wohland J, Perrette M, Golly A, Rogelj J, Childers K, Schewe J, Frieler K, Mengel M, Hare W, Schaeffer M (2016) Differential climate impacts for policy-relevant limits to global warming: the case of 1.5°C and 2°C. *Earth System Dynamics* 7(2):327-351. doi: 10.5194/esd-7-327-2016
- Smith HA, Fulton SE, McCleod IM, Page CA, Bourne DG (2023) Sea-weeding: Manual removal of macroalgae facilitates rapid coral recovery. *Journal of Applied Ecology* 60:2459-2471. doi: 10.1111/1365-2664.14502
- Sweatman H, Thompson A, Delean S, Davidson J, Neale S (2007) Status of near-shore reefs of the Great Barrier Reef 2004. Marine and Tropical Sciences Research Facility Research Report Series. Reef and Rainforest Research Centre Limited, Cairns 168pp
- Tanner JE (1995) Competition between scleractinian corals and macroalgae: an experimental investigation of coral growth, survival and reproduction. *Journal of Experimental Marine Biology and Ecology* 190(2):151-168. doi: 10.1016/0022-0981(95)00027-O
- Thompson A, Costello P, Davidson J (2015) Gladstone Healthy Harbour Partnership 2015 Report Card, ISP014: Coral. Australian Institute of Marine Science, Townsville. 41pp
- Thompson A, Davidson J, Logan M, Thompson C (2025) Marine Monitoring Program Annual Report for Inshore Coral Reef Monitoring: 2023–24. Report for the Great Barrier Reef Marine Park Authority, Great Barrier Reef Marine Park Authority, Townsville. 155 pp. <https://hdl.handle.net/11017/4124>
- Thompson C, Thompson A, Davidson J (2024) Coral Indicators for the 2024 Gladstone Harbour Report Card: ISP014. Report prepared for Gladstone Healthy Harbour Partnership. Australian Institute of Marine Science, Townsville. 46 pp.
- Van Dam JW, Negri AP, Uthicke S, Muller JF (2011) Chemical pollution on coral reefs: exposure and ecological effects. In: Sanchez-Bayo F, van den Brink PJ, Mann RM (Eds.), *Ecological Impact of Toxic Chemicals*. Bentham Science Publishers Ltd.
- Van Woesik R (1991) Immediate impact of the January 1991 floods on the coral assemblages of the Keppel Islands. Research Publication No. 23, Great Barrier Reef Marine Park Authority, Townsville
- Vega Thurber R, Burkepile DE, Correa AMS, Thurber AR, Shantz AA (2012) Macroalgae decrease growth and alter microbial community structure of the reef-building coral, *Porites astreoides*. *PLOS ONE* 7(9): e44246. doi: 10.1371/journal.pone.0044246
- Vision Environment (2013a) WBDDP 2013 Event Sampling –March 2013. Gladstone, QLD
- Vision Environment (2013b) WBDDP Water Quality Monitoring –April 2013. Gladstone, QLD
- Ward S, Harrison P, Hoegh-Guldberg O (2002) Coral bleaching reduces reproduction of scleractinian corals and increases susceptibility for future stress. Proc 9th International Coral Reef Symposium, Bali, Indonesia
- Waters DK, Carroll C, Ellis R, Hateley L, McCloskey GL, Packett R, Dougall C, Fentie B (2014) Modelling reductions of pollutant loads due to improved management practices in the Great Barrier Reef

catchments – Whole of GBR, Technical Report, Volume 1, Queensland Department of Natural Resources and Mines, Toowoomba, Queensland (ISBN: 978-1-7423-0999)

8 APPENDICES

8.1 Appendix 1: Data Tables

Table A 1 Site location and transect directions. Minor corrections from those detailed in Thompson *et al.* 2015 are included. Required maintenance of transect markers is indicated (*italic text*). At each transect a steel star picket marks the start point, then there are 10mm diameter sections of reinforcing bar at 10 m and at the end (20 m) of each transect. There is a 5 m gap between consecutive transects within each site.

Reef	Depth	Latitude	Longitude	Transect directions	
Seal Rocks North	1 m	-23°57.500	151°29.092	1	295 (end rod)
				2	285 then 310@10 m
				3	300 then 320@10 m
				4	30 then 105@10 m
				5	50 then 60@10 m
Seal Rocks South	1 m	-23°57.825	151°29.215	1	0 then 30@10 m
				2	30 then 350@10 m
				3	260 then 250@10 m (<i>10m rod</i>)
				4	190
				5	230 (<i>Picket, both rods</i>)
Rat Island	1 m	-23°46.022	151°19.107	1	305 then 300@10 m
				2	300 (<i>Picket, 10m rod</i>)
				3	330 then 320@10 m
				4	330 then 290@10 m
				5	285
Facing Island	0-1 m	-23°45.801	151°19.687	1	220 then 210@10 m
				2	190 then 180@10 m (<i>10m rod</i>)
				3	180 then 210@10 m
				4	240 then 230@10 m (end rod)
				5	170 (<i>Picket</i>)
Farmers Reef	1 m	-23°46.306	151°19.073	1	50
				2	40 then 50@10 m
				3	60
				4	60 then 75@10 m
				5	60 then 40@10 m (<i>end rod</i>)
Manning Reef	0-0.5	-23°51.239	151°21.199	1	30 then 10@10 m, 50 to T2
				2	60 then 0@10 m, 80 to T3
				3	60 then 300@10 m, 300 to T4
				4	315 then 20@10 m, 350 to T5
				5	320 then 40@10 m

Table A 2 Sub-indicator values for Gladstone Harbour. For the coral change sub-indicator the tabulated values are the mean of the changes in cover from the previous year, scores for this sub-indicator are based on a three-year rolling mean of these changes but only when reefs are not impacted by an acute disturbance event, and also consider the composition of the communities at each reef.

	Year	Juvenile density (m ²) *		Coral cover (%)		Cover change (%)		Macroalgae cover (%)	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD
Gladstone Harbour	2015	4.0	1.4	3.1	4.3	NA	NA	23.4	6.6
	2016	3.9	0.1	5.8	1.5	0.8	3.0	41.1	16.9
	2017	4.2	0.4	5.4	0.8	-0.4	2.3	35.3	24.9
	2018	3.7	0.6	4.2	0.7	-1.2	0.1	35.3	24.6
	2019	2.1	0.1	6.4	0.8	2.2	0.1	52.7	21.3
	2020	2.1	0.4	6.5	0.7	0.1	0.2	45.3	13.1
	2021	1.4	0.1	6.7	1.1	0.3	1.7	52.4	9.4
	2022	1.1	0.1	7.5	3.3	0.7	2.2	43.6	6.5
	2023	1.0	0.6	6.2	0.8	-1.2	2.4	55.5	21.5
	2024	1.0	0.5	6.4	0.8	0.2	0.0	43.9	6.2
	2025	0.9	0.0	4.4	1.1	-2.0	0.3	37.1	10.8

* Note: values given for juvenile densities are based on the current methodology and have been back calculated for previous years to allow comparison. This applies to all following tables of indicator values.

Table A 3 Indicator and sub-indicator scores for Gladstone Harbour.

	Year	Juvenile density	Coral cover	Coral change	Macroalgae cover	Coral Indicator	
						Score	Grade
Gladstone Harbour	2015	0.28	0.06	NA	0.19	0.18	E
	2016	0.34	0.07	NA	0.03	0.15	E
	2017	0.38	0.07	0.40	0.26	0.28	D
	2018	0.39	0.05	0.32	0.22	0.24	E
	2019	0.23	0.08	0.41	0.01	0.18	E
	2020	0.12	0.08	0.40	0.07	0.17	E
	2021	0.15	0.07	0.34	0.00	0.14	E
	2022	0.12	0.09	0.37	0.04	0.15	E
	2023	0.11	0.08	0.31	0.07	0.14	E
	2024	0.11	0.08	0.27	0.12	0.14	E
	2025	0.10	0.05	0.09	0.10	0.09	E

* Note: Juvenile density indicator scores are based on the current methodology and have been back calculated for previous years to allow comparison. Coral Indicator scores for previous years have also been adjusted accordingly. This applies to all following tables of indicator scores.

Table A 4 Sub-indicator values for reporting zones

Zone	Year	Juvenile density (m ⁻²)		Combined cover of hard and soft coral (%)		Change in hard coral cover (%)		Macroalgae cover (%)	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD
Mid Harbour	2016	3.9	0.5	4.7	3.1	-1.3	4.1	29.2	8.7
	2017	3.9	1.6	5.9	4.0	1.2	1.7	17.7	16.0
	2018	3.2	1.6	4.7	3.9	-1.2	2.0	17.9	14.9
	2019	2.2	0.7	7.1	6.0	2.3	2.3	37.7	19.8
	2020	2.3	1.4	7.0	4.5	-0.1	3.0	36.0	22.1
	2021	1.4	1.1	6.0	5.1	-0.9	1.9	45.8	16.2
	2022	1.2	0.9	5.2	3.8	-0.9	1.8	39.0	21.4
	2023	1.4	1.4	5.6	3.2	0.5	1.7	40.3	23.3
	2024	1.3	1.1	5.8	4.4	0.2	1.7	39.5	29.4
	2025	0.9	0.6	3.6	3.5	-2.2	3.4	44.8	31.0
Outer Harbour	2016	3.9	0.8	6.9	9.7	2.9	4.1	53.1	0.1
	2017	4.5	0.6	4.8	6.5	-2.0	3.2	52.9	7.6
	2018	4.1	0.4	3.8	4.4	-1.1	2.1	52.6	14.8
	2019	2.1	0.5	5.9	7.1	2.2	2.7	67.8	12.8
	2020	1.8	1.0	6.0	5.8	0.2	1.2	54.6	16.4
	2021	1.5	0.8	7.5	9.2	1.5	3.4	59.1	20.1
	2022	1.0	0.1	9.8	8.9	2.2	0.2	48.2	8.9
	2023	0.6	0.2	6.8	6.7	-2.9	2.0	70.7	11.8
	2024	0.6	0.2	6.9	6.3	0.2	0.6	48.2	7.9
	2025	1.0	0.2	5.1	4.6	-1.8	1.5	29.4	9.8

Table A 5 Indicator and Sub-indicator scores for reporting zones.

Zone	Year	Juvenile density	Coral cover	Coral change	Macroalgae cover	Coral Indicator	
						Score	Grade
Mid Harbour	2015	0.23	0.08		0.37	0.23	E
	2016	0.33	0.05		0.10	0.16	E
	2017	0.33	0.08	0.44	0.50	0.33	D
	2018	0.34	0.06	0.30	0.41	0.27	D
	2019	0.24	0.09	0.42	0.02	0.19	E
	2020	0.15	0.09	0.44	0.15	0.20	E
	2021	0.15	0.07	0.43	0	0.16	E
	2022	0.13	0.06	0.26	0.07	0.13	E
	2023	0.15	0.07	0.22	0.22	0.14	E
	2024	0.14	0.07	0.23	0.23	0.17	E
	2025	0.10	0.05	0.18	0.19	0.13	E
Outer Harbour	2015	0.33	0.05		0	0.13	E
	2016	0.33	0.09		0	0.14	E
	2017	0.44	0.06	0.37	0	0.21	E
	2018	0.45	0.05	0.33	0	0.20	E
	2019	0.22	0.07	0.40	0	0.17	E
	2020	0.08	0.08	0.39	0	0.14	E
	2021	0.15	0.07	0.26	0	0.12	E
	2022	0.11	0.12	0.48	0	0.18	E
	2023	0.06	0.08	0.40	0	0.14	E
	2024	0.07	0.09	0.32	0	0.12	E
	2025	0.10	0.06	0	0	0.04	E

Table A 6 Sub-indicator values for individual reefs. Values for change in coral cover are absolute change in hard coral cover between years.

Zone	Reef	Year	Juvenile density (m ²)	Coral cover (%)	Change in hard coral cover (%)	Macroalgae cover (%)
Mid Harbour	Facing Island	2015	5.0	13.1		24.8
		2016	4.3	6.1	-7.0	30.6
		2017	3.2	9.8	3.7	27.6
		2018	1.5	8.8	-1.0	14.5
		2019	1.5	13.5	4.8	40.5
		2020	0.6	9.1	-4.4	48.5
		2021	0.2	8.2	-0.9	63.2
		2022	0.6	5.3	-3.0	66
		2023	0.3	8.0	2.6	55.1
		2024	0.3	10.6	2.7	55.9
		2025	0.3	3.4	-7.3	72
	Farmers Reef	2015	3.2	4.8		4.1
		2016	3.8	7.1	2.7	35.9
		2017	6.3	7.2	0.0	5.8
		2018	5.2	3.0	-4.1	18.0
		2019	2.8	3.4	0.3	27.2
		2020	3.9	5.9	2.5	23.3
		2021	2.8	2.1	-3.6	32.2
		2022	2.5	3.6	1.1	16.5
		2023	3.4	3.6	0.4	13.0
		2024	2.8	3.1	-0.5	6.4
		2025	1.7	2.1	-1	9.2
	Manning Reef	2015	2.1	0.0		32.0
		2016	3.4	0.1	0.1	33.6
		2017	2.9	0.3	0.1	35.0
		2018	3.6	0.1	-0.1	37.5
		2019	1.6	0.6	0.5	64.2
		2020	1.8	1.2	0.6	60.1
		2021	1.6	1.4	0.1	55.8
		2022	0.9	1.5	0.1	44.8
		2023	0.5	2.1	0.6	63.6
		2024	0.8	1.2	-1.0	71.1
		2025	0.8	0.5	-0.8	69.4
	Rat Island	2015	1.8	6.4		14
		2016	4.3	5.5	-0.9	16.5
		2017	3.4	6.5	1.0	2.5
		2018	2.6	6.9	0.4	1.5
		2019	2.8	10.6	3.6	19.0
		2020	3.1	11.6	0.9	12.2

Mid Harbour	Rat Island	2021	0.9	12.1	0.7	31.8
		2022	0.7	10.4	-1.8	28.8
		2023	1.5	8.8	-1.6	29.6
		2024	1.3	8.4	-0.4	24.6
		2025	0.8	8.5	0.2	28.4

Zone	Reef	Year	Juvenile density (m ²)	Coral cover (%)	Change in hard coral cover (%)	Macroalgae cover (%)
Outer Harbour	Seal Rocks North	2015	5.0	0.0		28.0
		2016	4.4	0.0	0.0	53.0
		2017	4.1	0.2	0.3	58.2
		2018	3.9	0.6	0.4	63.1
		2019	1.7	0.9	0.3	76.8
		2020	1.1	1.9	1.0	66.2
		2021	0.9	1.0	-0.9	73.3
		2022	0.9	3.5	2.4	54.4
		2023	0.4	2.0	-1.5	79.1
		2024	0.8	2.5	0.6	53.8
		2025	1.1	1.9	-0.7	36.4
	Seal Rocks South	2015	3.2	8.3		58.2
		2016	3.3	13.8	5.8	53.1
		2017	5.0	9.4	-4.3	47.5
		2018	4.4	6.9	-2.6	42.1
		2019	2.4	10.9	4.1	58.8
		2020	2.5	10.1	-0.6	43.0
		2021	2.1	14.0	3.9	44.9
		2022	1.1	16.1	2.1	41.9
		2023	0.7	11.5	-4.4	62.4
		2024	0.5	11.4	-0.2	42.7
		2025	0.8	8.4	-2.9	22.5

Table A 7 Timeseries of indicator and sub-indicator scores for Mid Harbour reefs

Zone	Reef	Year	Scores					Grade
			Juvenile density	Coral cover	Coral change	Macroalgae cover	Coral Indicator	
Mid Harbour	Facing Island	2015	0.41	0.16		0	0.19	E
		2016	0.46	0.08		0	0.18	E
		2017	0.25	0.12	0.50	0	0.22	E
		2018	0.16	0.11	0.33	0.46	0.27	D
		2019	0.16	0.17	0.67	0	0.25	D
		2020	0	0.11	0.33	0	0.11	E
		2021	0.02	0.10	0.33	0	0.11	E
		2022	0.06	0.07	0	0	0.03	E
		2023	0.03	0.10	0.28	0	0.10	E
		2024	0.04	0.13	0.47	0	0.16	E
		2025	0.04	0.04	0.47	0	0.14	E
	Farmers Reef	2015	0.26	0.06		1.00	0.44	D
		2016	0.34	0.09		0	0.14	E
		2017	0.53	0.09	0.50	0.95	0.52	C
		2018	0.53	0.04	0.33	0.17	0.27	D
		2019	0.31	0.04	0.13	0	0.12	E
		2020	0.30	0.07	0.46	0	0.21	E
		2021	0.30	0.03	0.46	0	0.20	E
		2022	0.27	0.05	0.55	0.29	0.29	D
		2023	0.36	0.05	0.31	0.56	0.32	D
		2024	0.30	0.04	0.31	0.92	0.39	D
		2025	0.18	0.03	0.09	0.76	0.26	D
	Manning Reef	2015	0.12	0		0	0.04	E
		2016	0.25	0		0	0.08	E
		2017	0.22	0.01	0.51	0	0.18	E
		2018	0.40	0	0.27	0	0.17	E
		2019	0.17	0.01	0.29	0	0.12	E
		2020	0.08	0.02	0.34	0	0.11	E
		2021	0.18	0.02	0.38	0	0.14	E
		2022	0.10	0.02	0.24	0	0.09	E
		2023	0.05	0.03	0.19	0	0.07	E
		2024	0.09	0.02	0.15	0	0.06	E
		2025	0.09	0.01	0.10	0	0.06	E
	Rat Island	2015	0.11	0.08		0.50	0.23	E
		2016	0.39	0.07		0.29	0.25	D
		2017	0.31	0.08	0.24	1.00	0.41	D
		2018	0.28	0.09	0.26	1.00	0.41	D
		2019	0.31	0.14	0.59	0.09	0.28	D
		2020	0.22	0.15	0.31	0.60	0.32	D

Mid Harbour	Rat Island	2021	0.10	0.15	0.54	0	0.20	E
		2022	0.07	0.13	0.24	0	0.11	E
		2023	0.16	0.11	0.10	0	0.09	E
		2024	0.14	0.10	0	0	0.06	E
		2025	0.09	0.11	0.06	0	0.06	E

Table A 8 Timeseries of indicator and sub-indicator scores for Outer Harbour reefs.

Zone	Reef	Year	Scores					Grade
			Juvenile density	Coral cover	Coral change	Macroalgae cover	Coral indicator	
Outer Harbour	Seal Rocks North	2015	0.42	0		0	0.14	E
		2016	0.38	0		0	0.13	E
		2017	0.36	0.01	0.25	0	0.15	E
		2018	0.42	0.01	0.34	0	0.19	E
		2019	0.19	0.01	0.46	0	0.17	E
		2020	0.01	0.02	0.25	0	0.07	E
		2021	0.10	0.01	0.19	0	0.08	E
		2022	0.10	0.04	0.50	0	0.16	E
		2023	0.05	0.03	0.33	0	0.10	E
		2024	0.08	0.03	0.50	0	0.15	E
		2025	0.12	0.02	0	0	0.04	E
	Seal Rocks South	2015	0.25	0.10		0	0.12	E
		2016	0.28	0.17		0	0.15	E
		2017	0.51	0.12	0.50	0	0.28	D
		2018	0.48	0.09	0.33	0	0.22	E
		2019	0.26	0.14	0.33	0	0.18	E
		2020	0.15	0.13	0	0	0.07	E
		2021	0.20	0.12	0.33	0	0.16	E
		2022	0.12	0.20	0.47	0	0.20	E
		2023	0.07	0.14	0.47	0	0.17	E
		2024	0.05	0.14	0.14	0	0.08	E
		2025	0.09	0.10	0	0	0.05	E

Table A 9 Genus level coral cover and abundance of juvenile corals at reefs surveyed in 2025.

Sample Type	Location	Sample Type																			
		Acropora (Acroporidae)	Alveopora (Acroporidae)	Montipora (Acroporidae)	Duncanopsammia (Dendrophylliidae)	Turbinaria (Dendrophylliidae)	Leptastrea (Leptastreidae)	Micromussa (Lobophylliidae)	Moseleya (Lobophylliidae)	Coelastrea (Merulinidae)	Cyphastrea (Merulinidae)	Dipsastraea (Merulinidae)	Favites (Merulinidae)	Goniastrea (Merulinidae)	Paragoniastrea (Merulinidae)	Platygyra (Merulinidae)	Pocillopora (Pocilloporidae)	Bernardopora (Poritidae)	Goniopora (Poritidae)	Porites (Poritidae)	Psammocora (Psammocoridae)
Cover (%)	Facing Island	0	0	0	0	0.25	0	0	0	0	0.25	0	0	0	0	0	0	0	0	2.88	0
	Farmers Reef	0	0	0.38	0	0.50	0	0.13	0	0	0.75	0	0.25	0	0	0	0	0.13	0	0	0
	Manning Reef	0.25	0	0	0	0	0	0	0.13	0	0	0	0	0	0	0	0	0	0	0	0
	Rat Island	0.50	0	0	0	4.13	0.13	0	0	0	2.25	0	0	6.23	0	0	0	0	0	0.25	0
	Seal Rocks North	0.63	0	0.13	0	0.50	0	0	0	0	0	0	0	0	0	0	0.50	0	0	0	0
	Seal Rocks South	0.50	0	0.63	0	6.38	0	0	0	0	0	0	0	0	0	0	0.50	0	0	0	3.76
Juveniles (count)	Facing Island	3	0	2	0	1	0	0	0	0	1	0	0	0	0	0	0	0	0	3	0
	Farmers Reef	1	0	0	4	9	1	0	0	0	0	0	2	0	1	0	0	1	0	8	0
	Manning Reef	2	0	1	1	7	0	0	0	0	1	0	1	0	0	0	0	0	0	6	3
	Rat Island	0	0	0	0	1	1	0	0	0	3	0	0	0	0	0	0	0	0	10	1
	Seal Rocks North	4	1	0	0	10	0	0	1	0	0	1	1	0	3	0	1	0	1	0	5
	Seal Rocks South	3	0	0	0	4	0	0	0	0	0	1	3	0	2	1	0	0	0	0	1

Table A 10 Cover (%) of algae, sponges and sand and silt at reefs surveyed in 2025.

Location	Red Macroalgae			Brown Macroalgae					Green Macroalgae	Crustose coralline algae	Turf algae	Sand and Silt
	<i>Asparagopsis</i>	<i>Peyssonnelia</i>	Undefined	<i>Dictyota</i>	<i>Lobophora</i>	<i>Styopodium</i>	Sargassaceae	Undefined				
Facing Island	10	0	0.13	1.50	7.00	0	53.38	0	0	0.13	15.88	6.75
Farmers Reef	4.25	0	0.13	2.75	2.00	0	0	0.13	0	0	36.38	50.50
Manning Reef	43.50	0	0	6.88	19.00	0	0	0	0	0.13	8.75	21.13
Rat Island	22.63	0	0.25	4.13	1.38	0	0	0	0	0.75	24.25	35.25
Seal Rocks North	0	0.25	0.38	0.13	14.88	0.13	20.50	0.13	0	1.25	37.88	22.38
Seal Rocks South	0	0.50	0	0.13	9.00	0.63	12.00	0.25	0	0.25	30.13	38.25

Table A 11 Causes of coral mortality at time of survey. Area of survey 200 m2 at each reef. Data from 2019-2022 included for comparison. Bio-eroding sponge is primarily *Cliona orientalis*. Bleaching and physical damage are recorded as the proportion of colonies affected; a range indicates variability among transects.

Reef	Damage	Genus	Colonies affected						
			2019	2020	2021	2022	2023	2024	2025
Facing Island	Bio-eroding sponge	<i>Porites</i>	17	22	8	10	17	14	13
		<i>Turbinaria</i>			1				
	Atramentous necrosis	<i>Psammocora</i>					1	1	
	Bleaching			0-5%				1-5%	
Farmers Reef	Atramentous necrosis	<i>Cyphastrea</i>			2			1	1
	Bio-eroding sponge	<i>Cyphastrea</i>	5	7	4	7	8		6
		<i>Favites</i>							1
		<i>Plesiastrea</i>							1
		<i>Porites</i>				1		1	1
		<i>Turbinaria</i>	1						
	Bleaching			0-1%				1-5%	
Unknown	<i>Porites</i>				1				
Manning Reef	Bio-eroding sponge	<i>Psammocora</i>						1	1
	Bleaching								<1%
	Physical								<1%
	White syndrome	<i>Duncanopsammia</i>							1
Rat Island	Atramentous necrosis	<i>Cyphastrea</i>			1	7	1		
	Bio-eroding sponge	<i>Cyphastrea</i>	6	8	9	5	10	3	5
		<i>Plesiastrea</i>	2	1					
		<i>Porites</i>				1			
		<i>Turbinaria</i>	2	4	3	2	4	3	1
		<i>Favites</i>		1	1		1	1	1
	Black Band Disease	<i>Turbinaria</i>		1					
Bleaching			0-10%						
Seal Rocks North	Bleaching			1-50%			<1%	11-	
	Bio-eroding sponge	<i>Favites</i>					1		
		<i>Goniopora</i>					1		
		<i>Turbinaria</i>					3		
Seal Rocks South	Atramentous necrosis	<i>Turbinaria</i>			1				
	Bio-eroding sponge	<i>Turbinaria</i>	8	9	7	6	3	4	5
		<i>Goniopora</i>					1		
		<i>Favites</i>			1	1	1		
	Bleaching		0-1%	20-40%			<1%	6-30%	<1%
	Physical			<1%					<1%
Unknown	<i>Acropora</i>				1				

Table A 12 Size-class distribution of juvenile corals. Values are number of juveniles observed in 100m x 0.34m belt transects (34m²) at each reef. Data from all years of surveys included for comparison.

Reef	Size-class (cm)	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025
Facing Island	0-2	107	67	32	19	13	3	0	4	1	4	5
	2-5	28	58	58	20	27	15	5	12	6	5	5
Farmers Reef	0-2	32	47	64	56	30	24	21	18	18	17	9
	2-5	17	26	39	39	19	39	25	22	31	21	18
Manning Reef	0-2	52	55	49	46	18	18	8	7	6	7	10
	2-5	6	40	29	45	23	27	28	14	7	15	12
Rat Island	0-2	19	48	44	30	17	22	2	5	9	7	7
	2-5	23	43	28	26	44	33	14	7	20	18	9
Seal Rocks North	0-2	111	80	55	42	17	7	8	10	4	6	4
	2-5	31	48	64	69	36	23	19	14	8	14	24
Seal Rocks South	0-2	52	27	58	32	13	8	12	7	6	3	8
	2-5	30	55	58	64	44	39	30	13	11	7	7

8.2 Appendix 2: Rationale for sub-indicator selection and threshold setting.

8.2.1 Coral cover

For coral communities, the underlying assumption for resilience is that recruitment and subsequent growth of colonies is sufficient to compensate for losses resulting from the combination of acute disturbances and chronic adverse environmental conditions. High abundance of coral, expressed as proportional cover of the substratum, can be interpreted as an indication of resilience as the corals are clearly able to survive the ambient environmental conditions. In addition, high cover equates to a large brood-stock, a necessary link to recruitment and an indication of the potential for recovery of communities in the local area. Corals also contribute to the structural complexity of a reef and as such support increased biodiversity and provide important ecosystem services such as the provision of habitat for fishes. Finally, high cover is the most tangible reflection of a healthy coral community and a desirable state from an aesthetic perspective. The consideration of both hard and soft corals in this indicator recognises that all corals have a place on coral reefs and that the cover of an area by any coral is effectively mutually exclusive of another.

The selection of critical values or thresholds for coral cover about which to base assessments of condition is difficult. From MMP observations since 2005 there are no strong indications that either hard or soft coral cover varies substantially along water quality gradients suggesting a common Great Barrier Reef (GBR) wide threshold for coral cover is appropriate. We do, however, acknowledge that differing disturbance histories in space and time are likely to confound any analysis attempting to quantify such a relationship. For the MMP, the setting of a threshold for coral cover is still under discussion, however, is likely to be based on an aspirational target of ~50% cover. This target is informed by two prior assessments of coral cover on nearshore reefs. A broad scale survey of nearshore reefs between Cape Tribulation and the Keppel Islands using the same sampling methods as used in Gladstone Harbour undertaken in 2004 returned a mean cover of hard corals of 33% and of soft coral of 5% (Sweatman *et al.* 2007). This total coral cover mean of 38% was observed following the severe loss of corals that occurred as result of thermal bleaching in 1998 and also 2002 (Berkelmans *et al.* 2004) and so is considered too low as a threshold that would indicate “good condition”. Secondly, a summary of surveys from over 100 sites between Cape Flattery and the Keppel Islands prior to 1996 returned a mean cover of hard corals of 62% (Ayling 1997). In this second study, soft coral cover was not reported, and the surveys were based on a range of video and line intercept techniques. AIMS in-house analysis of coral cover estimates using line intercept (LIT) sampling along the same sites as photo point intercept (PIT) used by the MMP reveal a consistent bias with PIT being ~ 78% of that estimated by LIT ($r^2 = 0.99$). Correcting for technique puts the pre-1996 hard coral cover on inshore reefs at a mean of approximately 48%. Allowing some soft coral cover and rounding to an even percentage, the MMP is looking toward a threshold of 50% for the combined cover of hard and soft coral on inshore reefs. Finally, surveys conducted prior to 2009 in the Mid Harbour reporting zone of Gladstone Harbour had mean hard coral cover of 39% (BMT WBM 2013). Although the BMT WBM (2013) report did not provide a mean estimate for soft coral cover, Figure 4.4 of that report indicates soft coral cover in the Mid Harbour ranged between ~4% - 40%. These figures do not greatly deviate from the 50% combined cover of hard and soft corals likely to be used by the MMP in the future and so we suggest applying a 50% threshold for Gladstone also.

No prior data exist for the Outer Harbour reporting zone and so again we suggest a consistent use of the 50% threshold as this will allow comparison of condition across zones but also other regions of the GBR monitored by the MMP.

8.2.2 Macroalgae cover

Macroalgal (MA) recruitment, growth and biomass are controlled by a number of environmental factors such as the availability of suitable substratum, sufficient nutrients and light, and rates of herbivory (Schaffelke *et al.* 2005). High macroalgal abundance may suppress reef resilience (e.g., Hughes *et al.* 2007, Foster *et al.* 2008, Cheal *et al.* 2013; but see Bruno *et al.* 2009) by increasing competition for space or changing the microenvironment into which corals settle and grow (e.g., McCook *et al.* 2014, Hauri *et al.* 2010). On the GBR, high macroalgal cover correlates with high concentrations of chlorophyll, a proxy for nutrient availability (De'ath and Fabricius 2010). Once established, macroalgae pre-empt or compete with corals for space that might otherwise be available for coral growth or recruitment (e.g., Box and Mumby 2007, Hughes *et al.* 2007). For the purpose of this indicator, macroalgae are considered as species of the Rhodophyta (Red algae), Phaeophyta (Brown algae) and Chlorophyta (Green algae), excluding the encrusting coralline or short turf like species. The latter two groups are recorded as part of the assessments but are not aggregated into the MA indicator.

The interactions between corals and algae are complex, likely species-specific and, mostly, unquantified (McCook *et al.* 2014). Because of this it is difficult to determine realistic thresholds of macroalgal cover from which to infer information about the resilience of coral communities. Recent AIMS analysis of MMP data aimed at determining a threshold for the MA indicator gave a threshold of ~23% for communities in less than 3m depth below lowest astronomic tide (LAT), beyond which the density of juvenile corals declines. This direct influence on coral community replenishment could be used to define an upper bound for macroalgae cover. A further consideration is that within the MMP data set MA cover varies along environmental gradients with highest cover found in turbid areas and where wave or current action precludes the accumulation of fine sediments. As turbidity declines or the proportion of sediments with fine grain sizes increase then the cover of macroalgae also declines. This response to environmental conditions is a further constraint to the expectation of the level of MA cover at many locations. Current thinking within the MMP is to include the threshold mentioned above for an influence of juvenile corals as an upper threshold though reduce this to modelled estimates of cover based on observed relationships between MA cover, turbidity and sediment composition, in cases where these predictions are lower than the threshold for influence on juvenile corals. For the Gladstone Healthy Harbour Partnership monitoring, AIMS has collected sediment samples from each monitoring location and determined sediment grain size composition. The depth of these samples was only 1-2m below LAT and so will not be directly comparable to grain size compositions from MMP reefs that were sampled at the depth of 5m below LAT where wave driven resuspension is generally reduced. The results of the sediment analysis indicate that there is not a substantial accumulation of fine sediments at the coral sampling locations selected in Gladstone Harbour and this along with the limited depth of the reefs suggest turbidity and sedimentation will not be limiting macroalgae cover.

In light of the above considerations an upper bound of 20% cover of macroalgae was adopted for the Gladstone Harbour reefs as this is below the threshold for impacts to juvenile settlement at shallow depths but also recognises that macroalgae cover is a natural component of shallow reef communities in nearshore areas of the southern GBR. The most comparable reef monitored by AIMS to those in Gladstone Harbour is Pelican Island in Keppel Bay. At Pelican Island MA cover declined to ~5% as the coral community at 2m below LAT recovered. The lower bound for cover of MA on Gladstone Harbour reefs was set at 5% as this is in line with cover at Pelican Island during a period that corals were showing strong recovery from past disturbance events but also allowing some natural occurrence of MA. We suggest the threshold for cover for MA be set midway between the lower and upper bounds at 12.5%. We point out that the scoring of this indicator is the inverse to that used for coral cover or juvenile densities as high MA cover is considered a poor indication of coral community condition.

8.2.3 Juvenile density

Common disturbances to inshore reefs include cyclones (often associated with flooding), thermal bleaching, and outbreaks of crown-of-thorns seastar, all of which can result in widespread mortality of corals (e.g., Sweatman *et al.* 2007, Osborne *et al.* 2011). Recovery from such events is reliant on both the recruitment of new colonies and regeneration of existing colonies from remaining tissue fragments (Smith 2008, Diaz-Pulido *et al.* 2009). Previous studies have shown that elevated concentrations of nutrients, agrichemicals, and turbidity can negatively affect reproduction in corals (reviewed by Fabricius 2005, van Dam *et al.* 2011 Erftemeijer *et al.* 2012) and increased organic carbon concentrations can promote coral diseases and mortality (Kline *et al.* 2006, Kuntz *et al.* 2005). Furthermore, high rates of sediment deposition and accumulation on reef surfaces can affect larval settlement (Babcock and Smith 2002, Baird *et al.* 2003, Fabricius *et al.* 2003) and smother juvenile corals (Harrison and Wallace 1990, Rogers 1990, Fabricius and Wolanski 2000). Any of these water quality-related pressures on the early life stages of corals have the potential to suppress the resilience of communities reliant on recruitment for recovery. For these reasons the density of juvenile corals is an important indicator of coral community resilience, especially in periods following severe disturbance events.

The number of juvenile colonies observed along fixed area transects may be biased due to the different proportions of substratum available for coral recruitment. For example, live coral cover effectively reduces the space available for settlement of coral larvae, as do sandy or silty substrata onto which corals are unlikely or unable to settle. To create a comparative estimate of the density of juvenile colonies between reefs and through time, the numbers of recruits observed along fixed transects are converted to densities per area of transect that is 'available' for settlement. This standardisation divides the number of juvenile corals observed along fixed transects by the area of those fixed transects that is not occupied by existing corals or deposits of loose sediments to which corals could not settle.

The setting of a threshold against which to assess observed densities of juvenile corals is problematic as detailed demographic studies that allow the estimation of adequate levels of recruitment that are likely to ensure coral community resilience have not been undertaken for the range of communities present in the turbid nearshore waters of the GBR.

For the MMP selection of thresholds for the scoring of this metric was based on the analysis of recovery outcomes for MMP and LTMP reefs up to 2014 (Thompson *et al.* 2016), which provided a

baseline condition from which changes could be inferred as improvements or declines in condition. Changes to the methods for juvenile density estimates outline in this report requires thresholds are also adjusted. Previously, the thresholds for this metric were based on <10cm juvenile size classes, with a mean of 7.5 per m² of available substrate being the density at which the indicator score went from 'poor' to 'satisfactory'. For the revised estimates of juvenile corals (<5cm), the mean is 4.6 colonies m⁻², with the 10th and 90th percentiles of the distribution being 0 and 13 juveniles per m². These observations serve as a guide to the densities of juveniles that can be expected on inshore reefs.

One study that explicitly focused on estimating the density of juvenile corals (<10 cm) required for coral communities to recover rather than shift to an algal dominated state following severe disturbance suggested a threshold of 6.2 juveniles per m² (Graham *et al.* 2015). Because this work was undertaken in the Seychelles the relevance to the inshore GBR is unknown. However, considering the similarity between the inshore GBR mean and the threshold of Graham *et al.* 2015, the initial value of 7 juvenile colonies per m² of available substrate was adopted for the Gladstone Harbour threshold. As of 2018 a value of 4.6 will set the threshold to account for the reduced size class of <5cm and remains consistent with the threshold of Graham *et al.* 2015.

8.2.4 Cover change

This indicator metric is based on the rate at which hard coral cover increases. While high coral cover can justifiably be considered a positive indicator of community condition, the reverse is not necessarily true. Low cover may occur following acute disturbance and, hence, may not be a direct reflection of the community's resilience to underlying environmental conditions. For this reason, in addition to considering the actual level of coral cover we also assess the rate at which hard coral cover increases as a direct measure of recovery potential. This indicator reflects the coral growth performance on a per reef basis by comparing observed increase in coral growth (in the absence of acute disturbance) to expected coral growth. Estimates are derived by comparing the observed rate of change in hard coral cover at a given reef to that predicted by a multi-species form of the Gompertz growth equation (Dennis & Taper 1994, Ives *et al.* 2003). The equations used were parameterised from the time-series of coral cover from reefs monitored by the LTMP and the MMP over the period 1987-2007.

The growth models used are parameterised in a Bayesian framework to permit propagation of uncertainty from the two models onto the overall growth expected. For the Gladstone Harbour Report Card, the model is parameterised specifically for 2m depths. Observations of annual change in benthic cover derived from 47 near-shore reefs sampled over the period 1987-2007 were used to parameterise two multi-species Gompertz growth equations. These models returned estimates of growth rates for corals of the family Acroporidae and the combined grouping of all other hard corals. These two groups were modelled separately as the growth rate of Acroporidae is substantially higher than most other corals. Within these model's growth rate estimates are dependent on the cover of each of these hard-coral groups along with the cover of soft coral which in combination represent space competitors and so limit the area available for coral cover increase.

Model projections of future coral cover on GBR inshore reefs based on the growth rates estimated by these models coupled with the observed disturbance history for inshore reefs of the GBR over the period 1987-2002 indicated a long-term decline in coral cover (Thompson & Dolman 2010).

For this reason, the positive score of 1 was reserved for only those reefs at which the observed rate of change in cover exceeded twice the upper 95% confidence interval of the change predicted. Observations falling within the upper and lower confidence intervals of the change in predicted cover were scored as neutral (indicator score 0.5) and those not meeting the lower confidence interval of the predicted change received an indicator score of 0. The rate of change is averaged over three years of observations. As implemented in 2017 only two years of change were used (2015-2016 and 2017-2017), future applications will be based on a rolling mean of three years of observed changes. Years in which disturbance events occurred at particular reefs were not included as there is no logical expectation for an increase in cover in such situations.

8.2.5 References

- Ayling A (1997) The biological status of fringing reefs in the Great Barrier Reef world heritage area. Proceedings of the State of the Great Barrier Reef World Heritage Area Workshop pp. 109-113
- Babcock RC, Smith L (2002) Effects of sedimentation on coral settlement and survivorship. In: Proceedings of the 9th International Coral Reef Symposium, Bali, Indonesia, 245–248
- Baird AH, Babcock RC, Mundy CP (2003) Habitat selection by larvae influences the depth distribution of six common coral species. *Marine Ecology Progress Series* 252:289-293
- Berkelmans R, De'ath G, Kininmonth S, Skirving WJ (2004) A comparison of the 1998 and 2002 coral bleaching events on the Great Barrier Reef: spatial correlation, patterns and predictions. *Coral Reefs* 23:74-83
- BMT WBM (2013). Central Queensland corals and associated benthos: Monitoring review and gap analysis. April 2013. Prepared for the Gladstone Ports Corporation.
- Box SJ, Mumby PJ (2007) Effect of macroalgal competition on growth and survival of juvenile Caribbean corals. *Marine Ecology Progress Series* 342, 139-149.
- Bruno JF, Sweatman H, Precht WF, Selig ER, Schutte VGW (2009) Assessing evidence of phase shifts from coral to macroalgal dominance on coral reefs. *Ecology* 90:1478-1484
- Cheal AJ, Emslie M, MacNeil MA, Miller I, Sweatman H (2013) Spatial variation in the functional characteristics of herbivorous fish communities and the resilience of coral reefs. *Ecological Applications*, 23:174–188
- De'ath G, Fabricius KE (2010) Water quality as a regional driver of coral biodiversity and macroalgae on the Great Barrier Reef. *Ecological Applications* 20:840–850
- Dennis B, Taper ML (1994) Density dependence in time series observations of natural populations: estimation and testing. *Ecological Monographs* 64:205-224
- Diaz-Pulido G, McCook LJ, Dove S, Berkelmans R, Roff G, Kline DI, Weeks S, Evans RD, Williamson DH, Hoegh-Guldberg O (2009) Doom and boom on a resilient reef: Climate change, algal overgrowth and coral recovery. *PLoS ONE* 4(4): e5239
- Erftemeijer PLA, Reigl B, Hoeksema BW, Todd PA (2012) Environmental impacts of dredging and other sediment disturbances on corals: A review. *Marine Pollution Bulletin* 64:1737-1765
- Fabricius KE (2005) Effects of terrestrial runoff on the ecology of corals and coral reefs: Review and synthesis. *Marine Pollution Bulletin* 50:125-146
- Fabricius KE, Wild C, Wolanski E, Abele D (2003) Effects of transparent exopolymer particles and muddy terrigenous sediments on survival of hard coral recruits. *Estuarine, Coastal and Shelf Science* 57:613-621
- Fabricius KE, Wolanski E (2000) Rapid smothering of coral reef organisms by muddy marine snow. *Estuarine, Coastal and Shelf Science* 50:115-120
- Foster NL, Box SJ, Mumby PJ (2008) Competitive effects of macroalgae on the fecundity of the reef-building coral *Montastrea annularis*. *Marine Ecology Progress Series* 367:143-152

- Graham NAJ, Jennings S, MacNeil MA, Mouillot D, Wilson S (2015) Predicting climate-driven regime shifts versus rebound potential in coral reefs. *Nature* 518:94-97.
- Harrison PL, Wallace CC (1990) Reproduction, dispersal and recruitment of Scleractinian corals. In Dubinsky Z (ed) *Ecosystems of the world 25: Coral Reefs*. Elsevier, New York, pp 133-202
- Hauri C, Fabricius K, Schaffelke B, Humphrey C (2010) Chemical and physical environmental conditions underneath mat- and canopy-forming macroalgae, and the effects on understory corals. *PLoS ONE* 5(9): e12685
- Hughes TP, Rodrigues MJ, Bellwood DR, Ceccarelli D, Hoegh-Guldberg O, McCook L, Moltschaniwskyj N, Pratchett Ms, Steneck RS, Willis B (2007) Phase shifts, herbivory, and the resilience of coral reefs to climate change, *Current Biology* 17:360-365
- Ives AR, Dennis B, Cottingham KL, Carpenter SR (2003) Estimating community stability and ecological interactions from time series data. *Ecological Monographs* 73:301-330
- Kline DL, Kuntz NM, Breitbart M, Knowlton N, Rohwer F (2006) Role of elevated organic carbon levels and microbial activity in coral mortality. *Marine Ecology Progress Series* 314: 119–125
- Kuntz N, Kline D, Sandin S, Rohwer F (2005) Pathologies and mortality rates caused by organic carbon and nutrient stressors in three Caribbean coral species. *Marine Ecology Progress Series* 294:173–180
- Liu G, Heron S, Eakin C, Muller-Karger F, Vega-Rodriguez M, Guild L, De La Cour J, Geiger E, Skirving W, Burgess T, Strong A, Harris A, Maturi E, Ignatov A, Sapper J, Li J, Lynds S. 2014. Reef-scale thermal stress monitoring of coral ecosystems: new 5-km global products from NOAA coral reef watch. *Remote Sensing* 6(11):11579–11606 DOI 10.3390/rs6111579.
- McCook LJ, Jompa J, Diaz-Pulido G (2014) Competition between corals and algae on coral reefs: a review of evidence and mechanisms. *Coral Reefs* 19:400-417
- Osborne K, Dolman AM, Burgess SC, Johns KA (2011) Disturbance and the Dynamics of Coral Cover on the Great Barrier Reef (1995–2009). *PLoS ONE* 6:e17516
- Rogers CS (1990) Responses of coral reefs and reef organisms to sedimentation. *Marine Ecology progress Series* 62:185-202
- Schaffelke B, Mellors J, Duke NC (2005) Water quality in the Great Barrier Reef region: responses of mangrove, seagrass and macroalgal communities. *Marine Pollution Bulletin* 51:279-296
- Sweatman H, Thompson A, Delean S, Davidson J, Neale S (2007) Status of near-shore reefs of the Great Barrier Reef 2004. *Marine and Tropical Sciences Research Facility Research Report Series*. Reef and Rainforest Research Centre Limited, Cairns 168pp.
- Thompson A, Dolman A (2010) Coral bleaching: one disturbance too many for inshore reefs of the Great Barrier Reef. *Coral Reefs* 29:637-648
- Van Dam JW, Negri AP, Uthicke S, Muller JF (2011) Chemical pollution on coral reefs: exposure and ecological effects. In: Sanchez-Bayo F, van den Brink PJ, Mann RM (Eds.), *Ecological Impact of Toxic Chemicals*. Bentham Science Publishers Ltd.